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(54) Human serine protease

(57) Isolated nucleic acids encoding a human serine protease PSP1, protein obtainable from the nucleic acids, recombinant host cells transformed with the nucleic acids, oligonucleotides and primer pairs specific for *PSP1* polymorphisms and use of the protein and nucleic acid sequences are disclosed.

EP 0 828 003 A2

Description

The present invention relates to isolated human serine protease (PSP1) polynucleotides, their homologs and isoforms and polymorphic variants and their detection; to essentially pure PSP1 proteins; and to compositions and methods of producing and using PSP1 polynucleotides and proteins.

Mutations in the presenilins (PS-1 and PS-2) account for ~95% (75% and 20%, respectively) of all cases of early onset familial Alzheimer's disease (FAD). See R. Sherrington *et al., Nature 375,* 754-760 (1995); E.I. Rogaev *et al., Nature 376,* 775-778 (1995); and E. Levy-Lahad *et al., Science 269,* 973-977 (1995). The presenilins are highly homologous (67% identical), multi-membrane spanning proteins whose function is unknown.

It has been demonstrated that the 46 kDa full-length PS-1 protein is normally processed to 28 kDa and 18 kDa fragments; PS-2 has been reported to be similarly cleaved. See M. Mercken *et al., FEBS Letters 389,* 297-303 (1996). The predicted cleavage site(s) to account for fragments of this size would be in a region of the protein coded for by exon 8 and exon 9. Exon 8 is a hot spot for mutations leading to FAD. Thus, this region of PS-1, and potentially the cleavage of PS-1 in this region by a presenilinase protease, are important events in the functionality of the protein. A region of PS-1 spanning exons 8-11 has been demonstrated in the present invention to specifically bind a protease, PSP1, whose activity against its endogenous substrates and/or ability to bind to PS-1 are important in the pathology of neurodegeneration associated with AD, frontal lobe dementia, cortical lewy body disease, dementia of parkinson's disease, acute and chronic phases of degeneration following stroke or head injury, neuronal degeneration found in motor neurone disease, AIDS dementia and chronic epileps. Thus, a need exists for provision of the nucleotide and amino acid sequences corresponding to PSP1, for modulators of PSP1 binding to PS-1, and/or modulators of PSP1's proteolytic activity, for methods to identify such modulators and for reagents useful in such methods.

Accordingly, one aspect of the present invention is an isolated polynucleotide encoding a biologically active PSP1 polypeptide.

Another aspect of the invention is an isolated polynucleotide selected from the group consisting of:

(a) a polynucleotide encoding PSP1-1 having the nucleotide sequence as set forth in SEQ ID NO: 24 from nucleotide 603 to 1979; and
(b) a polynucleotide substantially similar to SEQ ID NO: 24.

Another aspect of the invention is an isolated polynucleotide selected from the group consisting of:

(a) a polynucleotide encoding PSP1-2 having the nucleotide sequence as set forth in SEQ ID NO: 23 from nucleotide 603 to 1979; and
(b) a polynucleotide substantially similar to SEQ ID NO: 23.

Another aspect of the invention is an isolated polynucleotide selected from the group consisting of:

(a) a polynucleotide encoding PSP1-3 having the nucleotide sequence as set forth in SEQ ID NO: 26 from nucleotide 603 to 1736; and
(b) a polynucleotide substantially similar to SEQ ID NO: 26.

Another aspect of the invention is an isolated polynucleotide selected from the group consisting of:

(a) a polynucleotide encoding PSP1-4 having the nucleotide sequence as set forth in SEQ ID NO: 28 from nucleotide 603 to 1913; and
(b) a polynucleotide substantially similar to SEQ ID NO: 28.

In a further aspect the invention provides any isolated polynucleotide as above defined wherein nucleotides 672 and 1435 are independently selected from C and T, hereinafter referred to as 'polymorphic variants'.

Another aspect of the invention is the functional polypeptides encoded by the polynucleotides of the invention.

Another aspect of the invention is an antisense oligonucleotide comprising a sequence which is capable of binding to the polynucleotides of the invention or D87258.

Another aspect of the invention is modulators of the polypeptides of the invention or of D87258.

Another aspect of the invention is a method for assaying a medium for the presence of a substance that modulates PSP1 or D87258 activity by affecting the binding of PSP1 or D87258 to cellular binding partners comprising the steps of:

(a) providing a PSP1 or D87258 protein having the amino acid sequence of PSP1-1, PSP1-2, PSP1-3 or PSP1-4 or D87258, or a functional derivative or polymorphic variant thereof and a cellular binding partner or synthetic

analog thereof;
(b) incubating with a test substance which is suspected of modulating PSP1 or D87258 activity under conditions which permit the formation of a PSP1 or D87258 protein/cellular binding partner complex;
(c) assaying for the presence of the complex, free PSP1 or D87258 protein or free cellular binding partner; and
(d) comparing to a control to determine the effect of the substance.

Another aspect of the invention is a method for assaying a medium for the presence of a substance that modulates PSP1 or D87258 activity by inhibiting proteolytic activity on a cellular substrate comprising the steps of:

(a) providing a PSP1 or D87258 protein having the amino acid sequence of PSP1-1, PSP1-2, PSP1-3 or PSP1-4 or D87258, or a functional fragment or polymorphic variant thereof and a cellular substrate or synthetic analog thereof;
(b) incubating with a test substance which is suspected of inhibiting PSP1 or D87258 activity under conditions which permit the formation of a PSP1 enzyme/substrate complex and subsequent cleavage of the substrate;
(c) assaying for the presence of proteolytically cleaved substrate; and
(d) comparing to a control to determine the effect of the substance.

Another aspect of the invention is a method for assaying for the presence of a substance that modulates PSP1 or D87258 activity by direct binding to PSP1 or D87258 protein comprising the steps of:

(a) providing a labelled PSP1 or D87258 protein having the amino acid sequence of PSP1-1, PSP1-2, PSP1-3 or PSP1-4 or D87258 or a functional derivative or polymorphic variant thereof;
(b) providing solid support-associated modulator candidates;
(c) incubating a mixture of the labelled PSP1 or D87258 protein with the support-associated modulator candidates under conditions which can permit the formation of a PSP1 protein/modulator candidate complex;
(d) separating the solid support from free soluble labelled PSP1 or D87258 protein;
(e) assaying for the presence of solid support-associated labelled protein;
(f) isolating the solid support complexed with labelled PSP1 or D87258 protein; and
(g) identifying the modulator candidate.

Another aspect of the invention is PSP1 or D87258 protein modulating compounds identified by the methods of the invention.

Another aspect of the invention is a method for the treatment of a patient having need to modulate PSP1 or D87258 activity comprising administering to the patient a therapeutically effective amount of the modulating compounds of the invention.

Another aspect of the invention is a method of diagnosing conditions associated with PSP1 or D87258 protein deficiency which comprises:

(a) isolating a polynucleotide sample from an individual;
(b) assaying the polynucleotide sample and a polynucleotide of the invention encoding PSP1 or D87258; and
(c) comparing differences between the polynucleotide sample and the PSP or D87258 polynucleotide, wherein any differences indicate mutations in the PSP1 or D87258 sequence.

Another aspect of the invention is a method of treating conditions which are related to insufficient PSP1 or D87258 protein function which comprises:

(a) isolating cells from a patient deficient in PSP1 or D87258 protein function;
(b) altering the cells by transfecting the polynucleotide of the invention or D87258 into the cells wherein a PSP1 or D87258 protein is expressed; and
(c) introducing the cells back to the patient to alleviate the condition.

Another aspect of the invention is a method of treating conditions which are related to insufficient PSP1 or D87258 protein function which comprises administering the polynucleotide of the invention to a patient deficient in PSP1 protein function wherein a PSP1 or D87258 protein is expressed and alleviates the condition.

Another aspect of the invention is an antibody immunoreactive with PSP1 or D87258 or an immunogen thereof.

Another aspect of the invention is a transgenic non-human animal capable of expressing in any cell thereof the polynucleotide of the invention.

Another aspect of the invention is a method for determining the genetic predisposition to neurodegeneration in a

patient comprising detecting PSP1 or D87258 polymorphisms in a sample from a patient. Yet another aspect of the invention is isolated polynucleotide having the nucleotide sequence as set forth in SEQ ID NO: 32, 33, 34, 35, 36 ,37, 38, 39, or 40.

Figure 1 is an amino acid sequence alignment of PSP1-1 with *E. coli* htrA.

Figure 2 is a multiple cDNA sequence alignment of the PSP1 isolates *PSP1-1, PSP1-2, PSP1-3* and *PSP1-4*.

Figure 3 is an amino acid sequence alignment of PSP1-1 with a putative human serine protease.

As used herein, the term "PSP1 polynucleotide" or *"PSP1"* refers to DNA molecules comprising a nucleotide sequence that encodes PSP1 and alternative splice variants, i.e., homologs and isoforms, and polymorphic variants. PSP1 binds to a region encompassing amino acids 269-413 of the human PS-1 protein, contains a conserved serine protease motif and exhibits homology to the *E. coli* serine protease htrA described by Lipinska *et al.* in *Nucl. Acids Res. 16,* 10053-10066 (1988) and a putative human serine protease with an IGF-binding motif (Ohno, I., *et al.,* Genbank Accession No. D87258 (1996)), hereinafter referred to as D87258.

The *PSP1-1* sequence is listed in SEQ ID NO: 24. The coding region of this sequence consists of nucleotides 603-1979 of SEQ ID NO: 24. The deduced 458 amino acid sequence of the encoded product PSP1-1 is listed in SEQ ID NO: 25.

The *PSP1-1* sequence listed in SEQ ID NO: 30 includes two polymorphic variants, at nucleotides 672 (C/T) and 1435 (C/T) resulting in alternative amino acid residues at position 24 (arg/cys) and 278 (ala/val), both in the conserved region of nucleotides 1-1540. The deduced 458 amino acid sequence of the encoded product *PSP1-1* is listed in SEQ ID NO: 31.

The *PSP1-2* sequence is listed in SEQ ID NO: 23. The coding region of this sequence consists of nucleotides 603-1979 of SEQ ID NO: 23. The deduced 458 amino acid sequence of the encoded product PSP1-2 is listed in SEQ ID NO: 8. The *PSP1-3* sequence is listed in SEQ ID NO: 26. The coding region of this sequence consists of nucleotides 603-1736 of SEQ ID NO: 26. The deduced 377 amino acid sequence of the encoded product PSP1-3 is listed in SEQ ID NO: 27. The *PSP1-4* sequence is listed in SEQ ID NO: 28. The coding region of this sequence consists of nucleotides 603-1913 of SEQ ID NO: 28. The deduced 436 amino acid sequence ofthe encoded product PSP1-4 is listed in SEQ ID NO: 29.

The D87258 sequence is listed in SEQ ID NO: 17. The coding region of this sequence consists of nucleotides 49-1491 of SEQ ID NO: 17. The deduced 480 amino acid sequence of the encoded product D87258 is listed in SEQ ID NO: 18. The D87258 sequence listed in SEQ ID NO: 17 includes a polymorphic variant at nucleotide 1325 (G/T) resulting in alternative amino acid residues at position 213 (gly/val). The sequence in Genbank Accession No. D87258 (1996)), describes only 1325G. The novel polynucleotide polymorph of D87258 having 1325T, is hereinafter referred to as D87258 (1325T) and the novel encoded product having valine at 213 is D87258 (1325T) protein. The novel polynucleotide D87258 (1325T) and its encoded protein can replace *PSP-1* in any of the composition, uses or methods herein described and such novel polypeptide, encoded protein, compositions, uses and methods also form part of the invention.

As used herein, the term "functional fragments" when used to modify a specific gene or gene product means a less than full length portion of the gene or gene product which retains substantially all of the biological function associated with the full length gene or gene product to which it relates. An example of a functional fragment of PSP1 is the minimal catalytic domain. To determine whether a fragment of a particular gene or gene product is a functional fragment, fragments are generated by well-known nucleolytic or proteolytic techniques or by the polymerase chain reaction and the fragments tested for the described biological function.

As used herein, an "antigen" refers to a molecule containing one or more epitopes that will stimulate a host's immune system to make a humoral and/or cellular antigen-specific response. The term is also used herein interchangeably with "immunogen."

As used herein, the term "epitope" refers to the site on an antigen or hapten to which a specific antibody molecule binds. The term is also used herein interchangeably with "antigenic determinant" or "antigenic determinant site."

As used herein, "monoclonal antibody" is understood to include antibodies derived from one species (e.g., murine, rabbit, goat, rat, human, etc.) as well as antibodies derived from two (or perhaps more) species (e.g., chimeric and humanized antibodies).

As used herein, a coding sequence is "operably linked to" another coding sequence when RNA polymerase will transcribe the two coding sequences into a single mRNA, which is then translated into a single polypeptide having amino acids derived from both coding sequences. The coding sequences need not be contiguous to one another so long as the expressed sequence is ultimately processed to produce the desired protein.

As used herein, "recombinant" polypeptides refer to polypeptides produced by recombinant DNA techniques; i.e., produced from cells transformed by an exogenous DNA construct encoding the desired polypeptide. "Synthetic" polypeptides are those prepared by chemical synthesis.

As used herein, a "replicon" is any genetic element (e.g., plasmid, chromosome, virus) that functions as an autonomous unit of DNA replication *in vivo;* i.e., capable of replication under its own control.

As used herein, a "vector" is a replicon, such as a plasmid, phage, or cosmid, to which another DNA segment may be attached so as to bring about the replication of the attached segment.

As used herein, a "reference" gene refers to the wild type PSP1 sequence of the invention and is understood to include the various sequence polymorphisms that exist, wherein nucleotide substitutions in the gene sequence exist, but do not affect the essential function of the gene product.

As used herein, a "mutant" gene refers to PSP1 sequences different from the reference gene wherein nucleotide substitutions and/or deletions and/or insertions result in perturbation of the essential function of the gene product.

As used herein, a DNA "coding sequence of' or a "nucleotide sequence encoding" a particular protein, is a DNA sequence which is transcribed and translated into a polypeptide when placed under the control of appropriate regulatory sequences.

As used herein, a "promoter sequence" is a DNA regulatory region capable of binding RNA polymerase in a cell and initiating transcription of a downstream (3' direction) coding sequence. For purposes of defining the present invention, the promoter sequence is bound at its 3' terminus by a translation start codon (e.g., ATG) of a coding sequence and extends upstream (5' direction) to include the minimum number of bases or elements necessary to initiate transcription at levels detectable above background. Within the promoter sequence will be found a transcription initiation site (conveniently defined by mapping with nuclease S1), as well as protein binding domains (consensus sequences) responsible for the binding of RNA polymerase. Eukaryotic promoters will often, but not always, contain "TATA" boxes and "CAT" boxes. Prokaryotic promoters contain Shine-Dalgarno sequences in addition to the -10 and -35 consensus sequences.

As used herein, DNA "control sequences" refers collectively to promoter sequences, ribosome binding sites, polyadenylation signals, transcription termination sequences, upstream regulatory domains, enhancers and the like, which collectively provide for the expression (i.e., the transcription and translation) of a coding sequence in a host cell.

As used herein, a control sequence "directs the expression" of a coding sequence in a cell when RNA polymerase will bind the promoter sequence and transcribe the coding sequence into mRNA, which is then translated into the polypeptide encoded by the coding sequence.

As used herein, a "host cell" is a cell which has been transformed or transfected, or is capable of transformation or transfection by an exogenous DNA sequence.

As used herein, a cell has been "transformed" by exogenous DNA when such exogenous DNA has been introduced inside the cell membrane. Exogenous DNA may or may not be integrated (covalently linked) into chromosomal DNA making up the genome of the cell. In prokaryotes and yeasts, for example, the exogenous DNA may be maintained on an episomal element, such as a plasmid. With respect to eukaryotic cells, a stably transformed or transfected cell is one in which the exogenous DNA has become integrated into the chromosome so that it is inherited by daughter cells through chromosome replication. This stability is demonstrated by the ability of the eukaryotic cell to establish cell lines or clones comprised of a population of daughter cells containing the exogenous DNA.

As used herein, "transfection" or "transfected" refers to a process by which cells take up foreign DNA and integrate that foreign DNA into their chromosome. Transfection can be accomplished, for example, by various techniques in which cells take up DNA (e.g., calcium phosphate precipitation, electroporation, assimilation of liposomes, etc.) or by infection, in which viruses are used to transfer DNA into cells.

As used herein, a "target cell" is a cell that is selectively transfected over other cell types (or cell lines).

As used herein, a "clone" is a population of cells derived from a single cell or common ancestor by mitosis. A "cell line" is a clone of a primary cell that is capable of stable growth *in vitro* for many generations.

As used herein, a "heterologous" region of a DNA construct is an identifiable segment of DNA within or attached to another DNA molecule that is not found in association with the other molecule in nature. Thus, when the heterologous region encodes a gene, the gene will usually be flanked by DNA that does not flank the gene in the genome of the source animal. Another example of a heterologous coding sequence is a construct where the coding sequence itself is not found in nature (e.g., synthetic sequences having codons different from the native gene). Allelic variation or naturally occurring mutational events do not give rise to a heterologous region of DNA, as used herein.

As used herein, a "modulator" of a polypeptide is a substance which can affect the polypeptide function, such as an inhibitor of enzymatic activity.

An aspect of the present invention is isolated polynucleotides encoding a PSP1 protein and substantially similar sequences. Isolated polynucleotide sequences are substantially similar if they are capable of hybridizing under moderately stringent conditions to SEQ ID NOs: 23, 24, 26 or 28 or they encode DNA sequences which are degenerate to SEQ ID NOs: 23, 24, 26 or 28 or are degenerate to those sequences capable of hybridizing under moderately stringent conditions to SEQ ID NOs: 23, 24, 26 or 28.

Moderately stringent conditions is a term understood by the skilled artisan and has been described in, for example, Sambrook *et al. Molecular Cloning: A Laboratory Manual,* 2nd edition, Vol. 1, pp. 101-104, Cold Spring Harbor Laboratory Press (1989). An exemplary hybridization protocol using moderately stringent conditions is as follows. Nitrocellulose filters are prehybridized at 65°C in a solution containing 6X SSPE, 5X Denhardt's solution (10g Ficoll, 10g BSA

and 10g polyvinylpyrrolidone per liter solution), 0.05% SDS and 100 ug/ml tRNA. Hybridization probes are labeled, preferably radiolabelled (e.g., using the Bios TAG-IT® kit). Hybridization is then carried out for approximately 18 hours at 65°C. The filters are then washed twice in a solution of 2X SSC and 0.5% SDS at room temperature for 15 minutes. Subsequently, the filters are washed at 58°C, air-dried and exposed to X-ray film overnight at -70°C with an intensifying screen.

Degenerate DNA sequences encode the same amino acid sequence as SEQ ID NOs: 8, 25, 27 or 29 or the proteins encoded by that sequence capable of hybridizing under moderately stringent conditions to SEQ ID NOs: 8, 25, 27, 29, but have variation(s) in the nucleotide coding sequences because of the degeneracy of the genetic code. For example, the degenerate codons UUC and UUU both code for the amino acid phenylalanine, whereas the four codons GGX, where X = U, C, A, or G, all code for glycine.

Alternatively, substantially similar sequences are defined as those nucleotide sequences encoding proteins having PSP1 activity in which about 70%, preferably about 80%, and most preferably about 90%, of the nucleotides share identity with *PSP1*, i.e., a sequence encoding a protein having PSP1 activity is substantially similar to any of SEQ ID NOs: 23, 24, 26 or 28 when at least about 70% of all of the nucleotides of the sequence match SEQ ID NOs: 23, 24, 26 or 28. Nucleotide sequences that are substantially similar can be identified by hybridization or by sequence comparison.

Embodiments of the isolated polynucleotides of the invention include DNA, genomic DNA and RNA, preferably of human origin. A method for isolating a nucleic acid molecule encoding a PSP1 protein is to probe a genomic or cDNA library with a natural or artificially designed probe using art recognized procedures. See, e.g., "Current Protocols in Molecular Biology", Ausubel *et al.* (eds.) Greene Publishing Association and John Wiley Interscience, New York, 1989,1992. The ordinarily skilled artisan will appreciate that SEQ ID NOs: 23, 24, 26 or 28 or fragments thereof comprising at least 15 contiguous nucleotides are particularly useful probes. It is also appreciated that such probes can be and are preferably labeled with an analytically detectable reagent to facilitate identification of the probe. Useful reagents include, but are not limited to, radioisotopes, fluorescent dyes or enzymes capable of catalyzing the formation of a detectable product. The probes would enable the ordinarily skilled artisan to isolate complementary copies of genomic DNA, cDNA or RNA polynucleotides encoding PSP1 proteins from human, mammalian or other animal sources or to screen such sources for related sequences, e.g., additional members of the family, type and/or subtype, including transcriptional regulatory and control elements as well as other stability, processing, translation and tissue specificity-determining regions from 5' and/or 3' regions relative to the coding sequences disclosed herein, all without undue experimentation.

Another aspect of the invention is functional polypeptides encoded by the polynucleotides of the invention and substantially similar polypeptides. An embodiment of a functional polypeptide of the invention is the PSP1 protein having the amino acid sequence set forth in SEQ ID NO: 8, 25, 27 or 29.

Polypeptide sequences that are substantially similar are those sequences having PSP activity in which about 50%, preferably 70%, and most preferably about 90%, of the amino acids share identity with PSPI, i.e., a sequence representing a polypeptide having PSP1 activity is substantially similar to any of SEQ ID NOs: 8, 24, 26 or 28 when at least about 50% of all of the amino acids of the sequence match SEQ ID NOs: 8, 25, 27 or 29. Substantially similar polypeptide sequences can be identified by techniques such as proteolytic digestion, gel electrophoresis, microsequencing and/or sequence comparison, e.g., through use of the GAP algorithm available from the University of Wisconsin Genetics Computer Group.

Another aspect of the invention is a method for preparing essentially pure PSP1 protein. Yet another aspect is the PSP1 protein produced by the preparation method of the invention. This protein has the amino acid sequence listed in SEQ ID NOs: 8, 25, 27 or 29 and includes variants with a substantially similar amino acid sequence that have the same function. The proteins of this invention are preferably made by recombinant genetic engineering techniques by culturing a recombinant host cell containing a vector encoding the polynucleotides of the invention under conditions promoting the expression of the protein and recovery thereof.

The isolated polynucleotides, particularly the DNAs, can be introduced into expression vectors by operatively linking the DNA to the necessary expression control regions, e.g., regulatory regions, required for gene expression. The vectors can be introduced into an appropriate host cell such as a prokaryotic, e.g., bacterial, or eukaryotic, e.g., yeast or mammalian cell by methods well known in the art. See Ausubel *et al., supra*. The coding sequences for the desired proteins, having been prepared or isolated, can be cloned into any suitable vector or replicon. Numerous cloning vectors are known to those of skill in the art and the selection of an appropriate cloning vector is a matter of choice. Examples of recombinant DNA vectors for cloning and host cells which they can transform include, but are not limited to, the bacteriophage *(E. coli)*, pBR322 *(E. coli)*, pACYC 177 *(E. coli)*, pKT230 (gram-negative bacteria), pGV1106 (gram-negative bacteria), pLAFR1 (gram-negative bacteria), pME290 (non-*E. coli* gram-negative bacteria), pHV14 *(E. coli* and *Bacillus subtilis)*, pBD9 *(Bacillus)*, pIJ61 *(Streptomyces)*, pUC6 *(Streptomyces)*, YIp5 *(Saccharomyces)*, a baculovirus insect cell system, a *Drosophila* insect system, YCp19 *(Saccharomyces)* and pSV2neo (mammalian cells). See generally, "DNA Cloning": Vols. I & II, Glover *et al.* ed. IRL Press Oxford (1985) (1987); and T. Maniatis *et al.* ("Molecular

Cloning" Cold Spring Harbor Laboratory (1982).

The gene can be placed under the control of control elements such as a promoter, ribosome binding site (for bacterial expression) and, optionally, an operator, so that the DNA sequence encoding the desired protein is transcribed into RNA in the host cell transformed by a vector containing the expression construct. The coding sequence may or may not contain a signal peptide or leader sequence. The proteins of the present invention can be expressed using, for example, the *E. coli* tac promoter or the protein A gene *(spa)* promoter and signal sequence. Leader sequences can be removed by the bacterial host in post-translational processing. See, e.g., U.S. Patent Nos. 4,431,739; 4,425,437 and 4,338,397.

In addition to control sequences, it may be desirable to add regulatory sequences which allow for regulation of the expression of the protein sequences relative to the growth of the host cell. Regulatory sequences are known to those of skill in the art. Exemplary are those which cause the expression of a gene to be turned on or off in response to a chemical or physical stimulus, including the presence of a regulatory compound or to various temperature or metabolic conditions. Other types of regulatory elements may also be present in the vector, for example, enhancer sequences.

An expression vector is constructed so that the particular coding sequence is located in the vector with the appropriate regulatory sequences, the positioning and orientation of the coding sequence with respect to the control sequences being such that the coding sequence is transcribed under the "control" of the control sequences, i.e., RNA polymerase which binds to the DNA molecule at the control sequences transcribes the coding sequence. Modification of the sequences encoding the particular antigen of interest may be desirable to achieve this end. For example, in some cases it may be necessary to modify the sequence so that it may be attached to the control sequences with the appropriate orientation; i.e., to maintain the reading frame. The control sequences and other regulatory sequences may be ligated to the coding sequence prior to insertion into a vector, such as the cloning vectors described above. Alternatively, the coding sequence can be cloned directly into an expression vector which already contains the control sequences and an appropriate restriction site.

In some cases, it may be desirable to produce mutants or analogues of PSP1 protein. Mutants or analogues may be prepared by the deletion of a portion of the sequence encoding the protein, by insertion of a sequence, and/or by substitution of one or more nucleotides within the sequence. Techniques for modifying nucleotide sequences, such as site-directed mutagenesis, are well known to those skilled in the art. See, e.g., T. Maniatis *et al., supra,* "DNA Cloning, " Vols. I and II, *supra;* and "Nucleic Acid Hybridization", *supra.*

Depending on the expression system and host selected, the proteins of the present invention are produced by growing host cells transformed by an expression vector described above under conditions whereby the protein of interest is expressed. Preferred mammalian cells include human embryonic kidney cells (293), monkey kidney cells, fibroblast (COS) cells, Chinese hamster ovary (CHO) cells, *Drosophila* or murine L-cells. If the expression system secretes the protein into growth media, the protein can be purified directly from the media. If the protein is not secreted, it is isolated from cell lysates or recovered from the cell membrane fraction. The selection of the appropriate growth conditions and recovery methods are within the skill of the art.

An alternative method to identify proteins of the present invention is by constructing gene libraries, using the resulting clones to transform *E. coli* and pooling and screening individual colonies using polyclonal serum or monoclonal antibodies to PSP1.

The proteins of the present invention may also be produced by chemical synthesis such as solid phase peptide synthesis on an automated peptide synthesizer, using known amino acid sequences or amino acid sequences derived from the DNA sequence of the genes of interest. Such methods are known to those skilled in the art.

The proteins of the present invention or their immunogenic fragments comprising at least one epitope can be used to produce antibodies, both polyclonal and monoclonal, directed to epitopes corresponding to amino acid sequences disclosed herein. If polyclonal antibodies are desired, a selected mammal such as a mouse, rabbit, goat or horse is immunized with a protein of the present invention, or its fragment, or a mutant protein. Serum from the immunized animal is collected and treated according to known procedures. Serum polyclonal antibodies can be purified by immunoaffinity chromatography or other known procedures.

Monoclonal antibodies to the proteins of the present invention, and to the immunogenic fragments thereof, can also be readily produced by one skilled in the art. The general methodology for making monoclonal antibodies by using hybridoma technology is well known. Immortal antibody-producing cell lines can be created by cell fusion and also by other techniques such as direct transformation of B lymphocytes with oncogenic DNA or transfection with Epstein-Barr virus. See, e.g., M. Schreier *et al.,* "Hybridoma Techniques" (1980); Hammerling *et al.,* "Monoclonal Antibodies and T-cell Hybridomas" (1981); Kennett *et al.,* "Monoclonal Antibodies" (1980); and U.S. Patent Nos. 4,341,761; 4,399,121; 4,427,783; 4,444,887; 4,452,570; 4,466,917; 4,472,500; 4,491,632; and 4,493,890. Panels of monoclonal antibodies produced against the antigen of interest, or fragment thereof, can be screened for various properties, i.e., for isotype, epitope, affinity, etc. Monoclonal antibodies are useful in purification, using immunoaffinity techniques, of the individual antigens which they are directed against. Alternatively, genes encoding the monoclonals of interest may be isolated from the hybridomas by PCR techniques known in the art and cloned and expressed in the appropriate vectors. The

antibodies of this invention, whether polyclonal or monoclonal have additional utility in that they may be employed as reagents in immunoassays, RIA, ELISA, and the like. The antibodies of the invention can be labeled with an analytically detectable reagent such as a radioisotope, fluorescent molecule or enzyme.

Chimeric antibodies, in which non-human variable regions are joined or fused to human constant regions (see, e. g., Liu *et al., Proc. Natl Acad. Sci. USA, 84,* 3439 (1987)), may also be used in assays or therapeutically. Preferably, a therapeutic monoclonal antibody would be "humanized" as described in Jones *et al., Nature, 321,* 522 (1986); Verhoeyen *et al., Science, 239,* 1534 (1988); Kabat *et al., J. Immunol., 147,* 1709 (1991); Queen *et al., Proc. Natl Acad. Sci. USA, 86,* 10029 (1989); Gorman *et al., Proc. Natl Acad. Sci. USA, 88,* 34181 (1991); and Hodgson *et al., Bio/Technology, 9:*, 421 (1991).

Another aspect of the present invention is modulators of the polypeptides of the invention or of D87258. Functional modulation of PSP1 or D87258 by a substance includes partial to complete inhibition of function, such as inhibition of proteolytic activity, identical function, as well as enhancement of function. Embodiments of modulators of the invention include peptides, oligonucleotides and small organic molecules including peptidomimetics. Modulators of the invention may be useful as therapeutics or prophylactics for all forms of neurodegeneration including AD. Modulators of PSP1 or D87258 proteolytic activity relative to other endogenous substrates may be also be useful for the treatment of other types of human disease states.

Another aspect of the invention is antisense oligonucleotides comprising a sequence which is capable of binding to the polynucleotides of the invention. Synthetic oligonucleotides or related antisense chemical structural analogs can be designed to recognize, specifically bind to and prevent transcription of a target nucleic acid encoding PSPI or D87258 protein by those of ordinary skill in the art. See generally, Cohen, J.S., *Trends in Pharm. Sci., 10,* 435(1989) and Weintraub, H.M., *Scientific American,* January (1990) at page 40.

Another aspect of the invention is a method for assaying a medium for the presence of a substance that modulates PSP1 or D87258 protein function by affecting the binding of PSP1 or D87258 protein to cellular binding partners. Examples of modulators include, but are not limited to peptides and small organic molecules including peptidomimetics. A PSP1 or D87258 protein is provided having the amino acid sequence of PSP1 (SEQ ID NOs: 8, 25, 27 or 29) or D87258 (SEQ ID NO: 18) or a functional fragment thereof together with a cellular binding partner or synthetic analog thereof. The mixture is incubated with a test substance which is suspected of modulating PSP1 or D87258 activity, under conditions which permit the formation of a PSP1 or D87258 gene product/cellular binding partner complex. An assay is performed for the presence of the complex, free PSP1 or D87258 protein or free cellular binding partner and the result compared to a control to determine the effect of the test substance.

Another aspect of the invention is a method for assaying a medium for the presence of a substance that modulates PSP1 or D87258 protein function by inhibiting its proteolytic activity on cellular substrates. Examples of modulators include, but are not limited to peptides and small organic molecules including peptidomimetics. Cellular substrates can include PS-1, PS-2, APP or other substrates. A PSP1 or D87258 protein is provided having the amino acid sequence of PSP1 (SEQ ID NOs: 8, 25, 27 or 29) or D87258 (SEQ ID NO: 18) or a functional fragment thereof together with a cellular substrate or synthetic analog thereof. The mixture is incubated with a test substance which is suspected of inhibiting PSP1 or D87258 activity, under conditions which permit the formation of a PSP1 or D87258 enzyme/substrate complex and subsequent cleavage of the substrate.

Another aspect of the invention is a method for assaying for the presence of a substance that modulates PSP1 or D87258 activity by direct binding to PSP1 or D87258 protein. Examples of modulators include, but are not limited to, peptides and small organic molecules including peptidomimetics. Modulator candidates are synthesized on a solid support by techniques such as those disclosed in Lam *et al., Nature 354,* 82 (1991) or Burbaum *et al., Proc. Natl. Acad. Sci. USA 92,* 6027 (1995) to provide solid support-associated modulator candidates. A labelled PSP1 or D87258 protein is provided having the amino acid sequence of PSP1 (SEQ ID NOs: 8, 25, 27 or 29) or D87258 (SEQ ID NO: 18) or a functional derivative thereof. Exemplary labels include directly attached fluorescent or colored dyes, biotin, radioisotopes or epitope tags, which are detectable by a suitable antibody. A mixture of solid support-associated modulator candidates and labelled PSP1 or D87258 protein is incubated under conditions which can permit the formation of a PSP1 or D87258 protein/modulator candidate complex. The solid support is separated from free soluble labelled PSP1 or D87258 protein. An assay is performed for the presence of solid support-associated labelled protein. Solid supports complexed with labelled protein are isolated and the identity of the modulator candidate determined by techniques well known to those skilled in the art, such as the TOF-SIMS method in Brummel *et al., Science* 264, 399-402 (1994).

Modulation of PSP1 or D87258 function would be expected to have effects on presenilin cleavage, the cleavage of other proteins or βA4 production. Any modulators so identified would be expected to be useful as a therapeutic for the treatment and prevention of neurodegeneration including FAD and AD.

Further, PSP1 or D87258 could be used to isolate proteins which interact with it and this interaction could be a target for interference. Inhibitors of protein-protein interactions between PSP1 or D87258 and other factors could lead to the development of pharmaceutical agents for the modulation of PSP1 or D87258 activity.

Methods to assay for protein-protein interactions, such as that of a PSP1 or D87258 gene product/binding partner

complex, and to isolate proteins interacting with PSP1 or D87258 are known to those skilled in the art. Use of the methods discussed below enable one of ordinary skill in the art to accomplish these aims without undue experimentation.

The yeast two-hybrid system provides methods for detecting the interaction between a first test protein and a second test protein, *in vivo,* using reconstitution of the activity of a transcriptional activator. The method is disclosed in U.S. Patent No. 5,283,173; reagents are available from Clontech and Stratagene. Briefly, PSP1 cDNA is fused to a *Gal4 or LexA* transcription factor DNA binding domain and expressed in yeast cells. cDNA library members obtained from cells of interest are fused to a transactivation domain of *Gal4* or another transactivation domain. cDNA clones which express proteins which can interact with PSP1 will lead to reconstitution of transcription factor activity such as *Gal4* and transactivation of a reporter gene expression such as *Gall-lacZ.*

An alternative method is screening of λgt11, λZAP (Stratagene) or equivalent cDNA expression libraries with recombinant PSP1. Recombinant PSP1 protein or fragments thereof are fused to small peptide tags such as FLAG, HSV or GST. The peptide tags can possess convenient phosphorylation sites for a kinase such as heart muscle creatine kinase or they can be biotinylated. Recombinant PSP1 can be phosphorylated with ^{32}[P] or used unlabeled and detected with streptavidin or antibodies against the tags. λgt11cDNA expression libraries are made from cells of interest and are incubated with the recombinant PSP1, washed and cDNA clones isolated which interact with PSP1. See, e. g., T. Maniatis *et al., supra.*

Another method is the screening of a mammalian expression library in which the cDNAs are cloned into a vector between a mammalian promoter and polyadenylation site and transiently transfected in COS or 293 cells followed by detection of the binding protein 48 hours later by incubation of fixed and washed cells with a labelled PSP1, prefereably iodinated, and detection of bound PSP1 by autoradiography (See Sims *et al., Science 241,* 585-589 (1988) and McMahan *et al., EMBO J. 10,* 2821-2832 (1991)). In this manner, pools of cDNAs containing the cDNA encoding the binding protein of interest can be selected and the cDNA of interest can be isolated by further subdivision of each pool followed by cycles of transient transfection, binding and autoradiography. Alternatively, the cDNA of interest can be isolated by transfecting the entire cDNA library into mammalian cells and panning the cells on a dish containing PSP1 bound to the plate. Cells which attach after washing are lysed and the plasmid DNA isolated, amplified in bacteria, and the cycle of transfection and panning repeated until a single cDNA clone is obtained (See Seed *et al, Proc. Natl. Acad. Sci. USA 84,* 3365 (1987) and Aruffo *et al., EMBO J.6,* 3313 (1987)). If the binding protein is secreted, its cDNA can be obtained by a similar pooling strategy once a binding or neutralizing assay has been established for assaying supernatants from transiently transfected cells. General methods for screening supernatants are disclosed in Wong *et al., Science 228,* 810-815 (1985).

Another alternative method is isolation of proteins interacting with PSP1 directly from cells. Fusion proteins of PSP1 with GST or small peptide tags are made and immobilized on beads. Biosynthetically labeled or unlabeled protein extracts from the cells of interest are prepared, incubated with the beads and washed with buffer. Proteins interacting with PSP1 are eluted specifically from the beads and analyzed by SDS-PAGE. Binding partner primary amino acid sequence data are obtained by microsequencing. Optionally, the cells can be treated with agents that induce a functional response such as tyrosine phosphorylation of cellular proteins. An example of such an agent would be a growth factor or cytokine such as erythropoietin or interleukin-3.

Another alternative method is immunoaffinity purification. Recombinant PSP1 is incubated with labeled or unlabeled cell extracts and immunoprecipitated with anti-PSP1 antibodies. The immunoprecipitate is recovered with protein A-Sepharose and analyzed by SDS-PAGE. Unlabelled proteins are labeled by biotinylation and detected on SDS gels with streptavidin. Binding partner proteins are analyzed by microsequencing. Further, standard biochemical purification steps known to those skilled in the art may be used prior to microsequencing.

Yet another alternative method is screening of peptide libraries for binding partners. Recombinant tagged or labeled PSP1 is used to select peptides from a peptide or phosphopeptide library which interact with PSP1. Sequencing of the peptides leads to identification of consensus peptide sequences which might be found in interacting proteins.

PSP1 or D87258 binding partners identified by any of these methods or other methods which would be known to those of ordinary skill in the art as well as those putative binding partners discussed above can be used in the assay method of the invention. Assaying for the presence of PSP1 or D87258 /binding partner complex are accomplished by, for example, the yeast two-hybrid system, ELISA or immunoassays using antibodies specific for the complex. In the presence of test substances which interrupt or inhibit formation of PSP1 or D87258 /binding partner interaction, a decreased amount of complex will be determined relative to a control lacking the test substance.

Assays for free PSP1 or D87258, or binding partner are accomplished by, for example, ELISA or immunoassay using specific antibodies or by incubation of radiolabeled PSP1 or D87258 with cells or cell membranes followed by centrifugation or filter separation steps. In the presence of test substances which interrupt or inhibit formation of PSP1 or D87258 /binding partner interaction, an increased amount of free PSP1 or D87258, or free binding partner will be determined relative to a control lacking the test substance.

Another aspect of the invention is pharmaceutical compositions comprising an effective amount of a PSP1 or

D87258 modulator of the invention and a pharmaceutically acceptable carrier. Pharmaceutical compositions of modulators of this invention for parenteral administration, i.e., subcutaneously, intramuscularly or intravenously or oral administration can be prepared.

The compositions for parenteral administration will commonly comprise a solution of the modulators of the invention or a cocktail thereof dissolved in an acceptable carrier, preferably an aqueous carrier. A variety of aqueous carriers may be employed, e.g., water, buffered water, 0.4% saline, 0.3% glycine and the like. These solutions are sterile and generally free of particulate matter. These solutions may be sterilized by conventional, well-known sterilization techniques. The compositions may contain pharmaceutically acceptable auxiliary substances as required to approximate physiological conditions such as pH adjusting and buffering agents, etc. The concentration of the modulator of the invention in such pharmaceutical formulation can vary widely, i.e., from less than about 0.5%, usually at or at least about 1% to as much as 15 or 20% by weight and will be selected primarily based on fluid volumes, viscosities, etc. according to the particular mode of administration selected.

Thus, a pharmaceutical composition of the modulator of the invention for intramuscular injection could be prepared to contain 1 mL sterile buffered water, and 50 mg of a protein of the invention. Similarly, a pharmaceutical composition of the modulator of the invention for intravenous infusion could be made up to contain 250 mL of sterile Ringer's solution, and 150 mg of a modulator of the invention. Actual methods for preparing parenterally administrable compositions are well known or will be apparent to those skilled in the art and are described in more detail in, for example, *Remington's Pharmaceutical Science,* 15th ed., Mack Publishing Company, Easton, Pennsylvania.

The physician will determine the dosage of the present therapeutic agents which will be most suitable and it will vary with the form of administration and the particular compound chosen, and furthermore, it will vary with the particular patient under treatment. Generally, the physician will wish to initiate treatment with small dosages substantially less than the optimum dose of the compound and increase the dosage by small increments until the optimum effect under the circumstances is reached. It will generally be found that when the composition is administered orally, larger quantities of the active agent will be required to produce the same effect as a smaller quantity given parenterally. The therapeutic dosage will generally be from 0.1 to 1000 milligrams per day and higher although it may be administered in several different dosage units.

Depending on the patient condition, the pharmaceutical composition of the invention can be administered for prophylactic and/or therapeutic treatments. In therapeutic applications, compositions containing the present compounds or a cocktail thereof are administered to a patient already suffering from a disease in an amount sufficient to cure or at least partially arrest the disease and its complications. In prophylactic applications, compositions containing the present compounds or a cocktail thereof are administered to a patient not already in a disease state to enhance the patient's resistance to the disease.

Single or multiple administrations of the pharmaceutical compositions can be carried out with dose levels and pattern being selected by the treating physician. In any event, the pharmaceutical composition of the invention should provide a quantity of the modulators of the invention sufficient to effectively treat the patient.

Additionally, some diseases result from inherited defective genes. These genes can be detected by comparing the sequence of the defective gene with that of a normal one. Individuals carrying mutations in the PSP1 or D87258 gene may be detected at the DNA level by a variety of techniques. Nucleic acids used for diagnosis (genomic DNA, mRNA, etc.) may be obtained from a patient's cells, such as from blood, urine, saliva or tissue biopsy, e.g., chorionic villi sampling or removal of amniotic fluid cells and autopsy material. The genomic DNA may be used directly for detection or may be amplified enzymatically by using PCR, ligase chain reaction (LCR), strand displacement amplification (SDA), etc. prior to analysis. See, e.g., Saiki *et al., Nature, 324,* 163-166 (1986), Bej, *et al., Crit. Rev. Biochem. Molec. Biol., 26,* 301-334 (1991), Birkenmeyer *et al., J. Virol. Meth., 35,* 117-126 (1991), Van Brunt, J., *Bio/Technology, 8,* 291-294 (1990)). RNA or cDNA may also be used for the same purpose. As an example, PCR primers complementary to the nucleic acid of the instant invention can be used to identify and analyze PSP1 or D87258 mutations. For example, deletions and insertions can be detected by a change in size of the amplified product in comparison to the normal PSP1 or D87258 genotype. Point mutations can be identified by hybridizing amplified DNA to rabiolabeled PSP1 or D87258 RNA of the invention or alternatively, radiolabelled PSP1 or D87258 antisense DNA sequences of the invention. Perfectly matched sequences can be distinguished from mismatched duplexes by RNase A digestion or by differences in melting temperatures (Tm). Such a diagnostic would be particularly useful for prenatal and even neonatal testing.

In addition, point mutations and other sequence differences between the reference gene and "mutant" genes can be identified by yet other well-known techniques, e.g., direct DNA sequencing, single-strand conformational polymorphism. See Orita *et al., Genomics, 5,* 874-879 (1989). For example, a sequencing primer is used with double-stranded PCR product or a single-stranded template molecule generated by a modified PCR. The sequence determination is performed by conventional procedures with radiolabeled nucleotides or by automatic sequencing procedures with fluorescent-tags. Cloned DNA segments may also be used as probes to detect specific DNA segments. The sensitivity of this method is greatly enhanced when combined with PCR. Further, point mutations and other sequence variations, such as polymorphisms, can be detected as described above, e.g., through the use of allele-specific oligonucleotides

for PCR amplification of sequences that differ by single nucleotides. Oligonucleotides having sequences as set forth in SEQ ID Nos: 32, 33, 34, 35, 36, 37, 38, 39 and 40 are useful in such a method. These methods are useful for determining the genetic predisposition to neurodegeneration in a patient by detecting polymorphisms within PSP1 or D87258 in a sample from a patient. Preferably, the polymorphisms detected are at nucleotide 672 of PSP1, at nucleotide 1435 of PSP1 or at nucleotide 1325 of D87258. Preferably, the polymorphisms are detected by PCR; most preferably, the polymorphisms are detected by PCR with oligonucleotides having a nucleotide sequence selected from the group consisting of SEQ ID NOs: 32, 33, 34, 35, 36, 37, 38, 39 and 40.Preferably, the neurodegeneration predisposition determined is to Alzheimer's disease.

Genetic testing based on DNA sequence differences may be achieved by detection of alteration in electrophoretic mobility of DNA fragments in gels with or without denaturing agents. Small sequence deletions and insertions can be visualized by high resolution gel electrophoresis. DNA fragments of different sequences may be distinguished on denaturing formamide gradient gels in which the mobilities of different DNA fragments are retarded in the gel at different positions according to their specific melting or partial melting temperatures. See, e.g., Myers *et al., Science, 230,* 1242 (1985). In addition, sequence alterations, in particular small deletions, may be detected as changes in the migration pattern of DNA heteroduplexes in non-denaturing gel electrophoresis such as heteroduplex electrophoresis. See, e. g., Nagamine *et al., Am. J. Hum. Genet., 45,* 337-339 (1989). Sequence changes at specific locations may also be revealed by nuclease protection assays, such as RNase and S1 protection or the chemical cleavage method as disclosed by Cotton *et al.* in *Proc. Natl. Acad. Sci. USA, 85,* 4397-4401 (1985).

Thus, the detection of a specific DNA sequence may be achieved by methods such as hybridization (e.g., heteroduplex electroporation, see, White *et al., Genomics, 12,* 301-306 (1992), RNAse protection (e.g., Myers *et al., Science, 230,* 1242 (1985)) chemical cleavage (e.g., Cotton *et al., Proc. Natl. Acad. Sci. USA, 85,* 4397-4401 (1985)), direct DNA sequencing, or the use of restriction enzymes (e.g., restriction fragment length polymorphisms (RFLP) in which variations in the number and size of restriction fragments can indicate insertions, deletions, presence of nucleotide repeats and any other mutation which creates or destroys an endonuclease restriction sequence). Southen blotting of genomic DNA may also be used to identify large (i.e., greater than 100 base pair) deletions and insertions.

In addition to conventional gel electrophoresis and DNA sequencing, mutations such as microdeletions, aneuploidies, translocations, inversions, can also be detected by *in situ* analysis. See, e.g., Keller *et al.,* DNA Probes, 2nd Ed., Stockton Press, New York, N.Y., USA (1993). That is, DNA or RNA sequences in cells can be analyzed for mutations without isolation and/or immobilization onto a membrane. Fluorescence *in situ* hybridization (FISH) is presently the most commonly applied method and numerous reviews of FISH have appeared. See, e.g., Trachuck *et al., Science, 250,* 559-562 (1990), and Trask *et al., Trends, Genet., 7,* 149-154 (1991). Hence, by using nucleic acids based on the structure of the PSPI or D87258 genes, one can develop diagnostic tests for genetic mutations.

In addition, some diseases are a result of, or are characterized by, changes in gene expression which can be detected by changes in the mRNA. Alternatively, the PSP1 or D87258 gene can be used as a reference to identify individuals expressing an increased or decreased level of PSP1 or D87258 mRNA, e.g., by Northern blotting or *in situ* hybridization.

Defining appropriate hybridization conditions is within the skill of the art. *See, e.g.,* "Current Protocols in Mol. Biol. " Vol. I & II, Wiley Interscience. Ausbel *et al.* (eds.) (1992). Probing technology is well known in the art and it is appreciated that the size of the probes can vary widely but it is preferred that the probe be at least 15 nucleotides in length. It is also appreciated that such probes can be and are preferably labeled with an analytically detectable reagent to facilitate identification of the probe. Useful reagents include but are not limited to radioisotopes, fluorescent dyes or enzymes capable of catalyzing the formation of a detectable product. As a general rule, the more stringent the hybridization conditions the more closely related genes will be that are recovered.

The putative role of PSP1 or D87258 in presenilin biochemistry establishes yet another aspect of the invention which is gene therapy. "Gene therapy" means gene supplementation where an additional reference copy of a gene of interest is inserted into a patient's cells. As a result, the protein encoded by the reference gene corrects the defect and permits the cells to function normally, thus alleviating disease symptoms. The reference copy would be a wild-type form of the *PSP1* or *D87258* gene or a gene encoding a protein or peptide which modulates the activity of the endogenous PSP1 or D87258.

Gene therapy of the present invention can occur *in vivo* or *ex vivo. Ex vivo* gene therapy requires the isolation and purification of patient cells, the introduction of a therapeutic gene and introduction of the genetically altered cells back into the patient. A replication-deficient virus such as a modified retrovirus can be used to introduce the therapeutic *PSP1* or *D87258* gene into such cells. For example, mouse Moloney leukemia virus (MMLV) is a well-known vector in clinical gene therapy trials. See, e.g., Boris-Lauerie *et al., Curr. Opin. Genet. Dev., 3,* 102-109 (1993).

In contrast, *in vivo* gene therapy does not require isolation and purification of a patient's cells. The therapeutic gene is typically "packaged" for administration to a patient such as in liposomes or in a replication-deficient virus such as adenovirus as described by Berkner, K.L., in *Curr. Top. Microbiol. Immunol., 158,* 39-66 (1992) or adeno-associated virus (AAV) vectors as described by Muzyczka, N., in *Curr. Top. Microbiol. Immunol., 158,* 97-129 (1992) and U.S.

Patent No. 5,252,479. Another approach is administration of "naked DNA" in which the therapeutic gene is directly injected into the bloodstream or muscle tissue. Another approach is administration of "naked DNA" in which the therapeutic gene is introduced into the target tissue by microparticle bombardment using gold particles coated with the DNA.

Cell types useful for gene therapy of the present invention include lymphocytes, hepatocytes, myoblasts, fibroblasts, any cell of the eye such as retinal cells, epithelial and endothelial cells. Preferably the cells are T lymphocytes drawn from the patient to be treated, hepatocytes, any cell of the eye or respiratory or pulmonary epithelial cells. Transfection of pulmonary epithelial cells can occur via inhalation of a neubulized preparation of DNA vectors in liposomes, DNA-protein complexes or replication-deficient adenoviruses. See, e.g., U.S. Patent No. 5,240,846.

Another aspect of the invention is transgenic, non-human mammals capable of expressing the polynucleotides of the invention or D87258 in any cell. Transgenic, non-human animals may be obtained by transfecting appropriate fertilized eggs or embryos of a host with the polynucleotides of the invention, with D87258 or with mutant forms found in human diseases. See, e.g., U.S. Patent Nos. 4,736,866; 5,175,385; 5,175,384 and 5,175,386. The resultant transgenic animal may be used as a model for the study of *PSP1* or *D87258* gene function. Particularly useful transgenic animals are those which display a detectable phenotype associated with the expression of the PSP1 or D87258 protein. Drug development candidates may then be screened for their ability to reverse or exacerbate the relevant phenotype.

The present invention will now be described with reference to the following specific, non-limiting examples.

Example 1 - Identification of the PS-1 Binding Partner PSP1

A portion of PS-1 cDNA (GenBank Accession No. L42110) (SEQ ID NO: 9) encoding residues 269-413 of the PS-1 amino acid sequence (SEQ ID NO: 10) was PCR amplified with the oligonucleotide primers 5'-CGGAATTCCGTATGCTGGTTGAAACA-3' (SEQ ID NO: 11) and 5'-CGGGATCCTCAGGCTACGAAACAGGCTAT-3' (SEQ ID NO: 12). The product was digested with *Eco*RI and *BamHI* and cloned into pEG202 (Golemis *et al.*, in *Current Protocols in Molecular Biology*, John Wiley & Sons, New York (1994)). The resulting plasmid, pCC352, encoded a fusion protein in which the DNA binding protein, *LexA*, was fused in-frame to amino acids 269-413 of PS-1. The parent vector, pEG202, was a yeast expression vector which uses the alcohol dehydrogenase *(ADH1)* promoter to express the *LexA* fusion proteins and *HIS3* as the selectable marker. Sequence analysis using an automated DNA sequencer (Applied Biosystems, Inc.) confirmed that the amplified region had the correct sequence and was fused in-frame to *LexA*.

All procedures, plasmids and strains used in the two-hybrid screen have been described in detail by Golemis *et al., supra.* Yeast strain EGY48 *(MATα, trp1, his3, ura3, 6ops-LEU2)* was cotransformed with the plasmids pCC352 and pSH18-34. Transformants were selected using complete minimal media lacking uracil and histidine. The plasmid pSH18-34 is a yeast expression vector in which eight *LexA* operator sites are located upstream of a minimal *GAL1* promoter which drives the expression of the *LacZ* gene and *URA3* as a selectable marker. Synthesis of the full length LexA-PS-1 fusion was confirmed by Western blot analysis of yeast extracts using polyclonal antisera directed against LexA. It was confirmed that the LexA-PS-1 fusion alone was unable to activate neither the *LEU2* nor *LacZ* reporter strains. In addition, the ability of the LexA-PS-1 fusion to enter the nucleus and bind DNA was confirmed using a repression assay.

A strain containing the LexA-PS-1 fusion and pSH18-34 (CCY321) was transformed with a human fetal brain cDNA library (Clontech) in plasmid pJG4-5 using a library scale transformation protocol. This library plasmid contains the *TRP1* selectable marker and allows the expression of cDNAs as fusions (AD fusions) to a cassette containing the SV40 nuclear localization sequence, the acid blob B42, and the hemagglutinin epitope tag. See Gyuris *et al., Cell 75,* 791-803 (1993). Expression of this fusion is under control of the galactose inducible promoter *GAL1*. Transformation reactions were plated onto complete minimal media lacking uracil, histidine and tryptophan. Approximately 4.5×10^6 individual transformants were obtained, pooled and frozen. To ensure that each primary colony was replated during the selection procedure, 2×10^7 viable cells (approximately 3 times the number of individual transformants) were plated onto minimal media lacking uracil, histidine, tryptophan and leucine with galactose/raffinose as the carbon source to induce expression of AD fusions. Colonies arising after 3 and 4 days of growth at 30 °C were picked to complete minimal media lacking uracil, histidine and tryptophan. Colonies containing potential interacting fusion proteins were then tested for galactose dependence and LacZ expression. Those isolates which activated both the *LEU2* and *LacZ* reporters in a galactose dependent fashion were considered positive and pursued further. Plasmids were isolated from yeast, used to transform *E. coli* strain KC8, and AD fusion plasmids selected by growth on minimal *E. coli* media lacking tryptophan. Each AD fusion plasmid containing a potential interacting fusion was used to transform CCY321. Several transformants were subjected to screening for galactose dependent *LEU2* and *LacZ* activation. To ensure that the interaction was specific, the ability of each AD fusion plasmid to interact with 22 nonrelated *LexA* fusion proteins was tested. AD fusion plasmids which passed this second round of screening and interacted specifically with the LexA-PS-1 fusion were identified.

Example 2 - *PSP1* cDNA Cloning and Sequence Analysis

The AD fusion plasmids were subjected to restriction digest analysis and sequencing as indicated above. Sequence analysis of one of the interacting fusion protein cDNAs revealed a 519 nucleotide open reading frame (SEQ ID NO: 1) encoding a 173 amino acid (SEQ ID NO: 2) protein starting with an GGA at position 2 and terminating with a TGA at position 523 of SEQ ID NO: 1. GenBank searches using the BLASTX and BLASTN algorithms with the cDNA sequence or with the deduced amino acid sequence indicated homology to a portion of the *E. coli* serine protease htrA described by Lipinska *et al., supra,* (SEQ ID NOs: 13 and 14). This novel cDNA was designated PSP1.

To obtain a greater portion of the cDNA, the oligonucleotide, 5'-CTGGATGGGGAGGTGATTGGAGTG-3' (SEQ ID NO: 15) representing bp 83-106 of SEQ ID NO: 1, was used to screen a Superscript human brain cDNA library (Gibco BRL) using the Genetrapper cDNA positive selection system (Gibco BRL). Colonies were screened using whole cell PCR or standard hybridization conditions as described by Innis *et al., PCR Protocols: A Guide to Methods and Applications,* Academic Press, San Diego, CA (1990) and Sambrook *et al., Molecular Cloning: A Laboratory Manual,* 2nd ed., Cold Spring Harbor Press, Cold Spring Harbor, NY (1989). Those isolates which contained PSPI were subjected to restriction digest analysis and sequencing. The longest clones, SEQ ID NO: 3 and SEQ ID NO: 5 were sequenced in their entirety.

Sequence analysis of SEQ ID NO: 3 revealed a 969 nucleotide open reading frame encoding a 323 amino acid (SEQ ID NO: 4) protein starting with a CCC at position 1 and terminating with a TGA at position 972 of SEQ ID NO: 3. Sequence analysis of SEQ ID NO: 5 revealed a 1500 nucleotide open reading frame encoding a 423 amino acid (SEQ ID NO: 6) protein starting with an CTT at position 1 and terminating with a TGA at position 1272 of SEQ ID NO: 5.

A second round of screening was performed using the oligonucleotide, 5'-GTCTCTGGGCCCCGGTTGTCTGTTG-3' (SEQ ID NO: 16) representing bp 5-28 of SEQ ID NO: 5; the library and screening protocol remained unchanged. In the second round of screening, the isolate designated SEQ ID NO: 7 contained the longest cDNA clone. Sequence analysis of SEQ ID NO: 7 revealed a 1374 nucleotide open reading frame encoding a 458 amino acid (SEQ ID NO: 8) protein starting with an ATG at position 251 and terminating with a TGA at position 1627 of SEQ ID NO: 7. However, SEQ ID NO: 7 does not have a stop codon upstream from the potential initiation codon. To confirm that the predicted start codon is authentic, the 5' nucleotide sequence was extended with 5' RACE using "Marathon Ready" human brain cDNA (Clontech) and a nested set of primers. A SEQ ID NO: 7 specific primer 5'-CCAACAGACAACCGGGCCCAGA-GACT-3' (SEQ ID NO: 20) and a 5' anchor primer-1 (Clontech) was used in the first PCR amplification and a SEQ ID NO: 7 specific primer 5'-TGCCTCCTCGCCCGCCCTACTCAGA-3' (SEQ ID NO: 21) and 5' anchor primer-2 (Clontech) was used in the second PCR amplification. PCR products were T/A cloned into pCR2.1 (Invitrogen). Eighteen isolates with staggered 5' ends were analyzed and a 5' consensus sequence of 587 nucleotides was generated (SEQ ID NO: 22). Alignment of SEQ ID NO: 22 and SEQ ID NO:7 to generate a consensus sequence (SEQ ID NO: 23) indicates that at nucleotide position 225 there is an in frame stop codon and the first methionine corresponds to that predicted in SEQ ID NO: 7. This gene is designated *PSP1-2.*

Consensus full length sequences for the genes designated *PSP1-1* (SEQ ID NOs: 24 and 25), *PSP1-3* (SEQ ID NOs: 26 and 27) and *PSP1-4* (SEQ ID NOs: 28 and 29) were generated from alignment of the 5' consensus sequence (SEQ ID NO: 22), other partial PSP1 clones, and with SEQ ID NOs: 7, 3 and 5, respectively.

Alignment of the deduced amino acid sequence of *PSP1-1* (SEQ ID NO: 25) to *E. coli* htrA (SEQ ID NO: 14) was accomplished using the BESTFIT algorithm (University of Wisconsin Genetics Computer Group). An approximate similarity of 55% and an identity of 33.5% at the amino acid level was observed and is shown in Fig. 1 (top, *PSP1-1;* bottom, *E. coli htrA).* The critical histidine and serine motif GXSXG conserved in all serine proteases is present in PSP1-1 at amino acid positions 198 and 304-308, respectively, and are indicated in bold. Amino acid numbers are indicated at the left and right of the sequence alignment.

Nucleotide sequence comparison of *PSP1-2, PSP1-1, PSP1-3* and *PSP1-4* using the PILEUP and PRETTY algorithms (University of Wisconsin Genetics Computer Group) with gap creation and extension penalties of 5.0 and 0.3, respectively, is shown in Fig. 2. The alignment results indicate that at nucleotide position 1541 of the alignment, *PSP1-2* and *PSP1-1* contain a 225 bp deletion and *PSP1-4* contains a 195 bp deletion. Within the same alignment at nucleotide position 1942, *PSP1-4* lacks 96 bp that are present in *PSP1-2, PSP1-1 and PSP1-3.* At the junction of each deletion site there is a splice site consensus sequence AGG or TGG (indicated in bold), suggesting that these alternate forms are due to alternative splicing. *See* Mount,S. in *Nucl. Acids Res 10,* 458-472 (1982). The apparent splicing event at position 1541 results in the removal of a stop codon (underlined in Fig. 2) that is present in *PSP1-3.* In addition, *PSP1-2* and *PSP1-1* contain a single nucleotide difference at position 672 of the alignment. *PSP1-2* contains a T at this position producing the codon TGC which codes for a cysteine while *PSP1-1* contains a C at the same position producing the codon CGC which codes for a cysteine.

Nucleotide sequence comparison of *PSP1-1* (SEQ ID NO: 24) to the putative human serine protease of Ohno *et al., supra,* (SEQ ID NO: 17) indicated a 49% identity using the GAP algorithm and 65% using the BESTFIT algorithm (data not shown). Alignment of the deduced amino acid sequence of *PSP1-1* (SEQ ID NO: 25) to the D87258 protease

of Ohno *et al., supra,* (SEQ ID NO: 18) was accomplished using the BESTFIT algorithm and is shown in Fig. 3 (top, PSP1-1; bottom, Ohno *et al.* D87258 protease). An approximate identity of 46% at the amino acid level was observed.

Example 3 - Tissue Distribution of PSP1

Northern analysis was carried out to determine the distribution of *PSP1* mRNA in human tissues. A 30-base oligonucleotide probe directed against the *PSP1* sequence was used (5'-ATGCTGAACATCGGGAAAGCTTGGTTCTCG-3') (SEQ ID NO: 19). This probe was 3'-end labelled with [^{32}P]-dATP. Northern blots containing mRNA from multiple human tissues (Clontech #7750-1, #7760-1, and #7755-1) were hybridized with this probe under stringent conditions. A major band of approximately 1.9kb was detected in all regions investigated: heart, brain, lung, placenta, liver, skeletal muscle, kidney, pancreas, amygdala, caudate nucleus, corpus callosum, hippocampus, substantia nigra, subthalamic nucleus, thalamus, cerebellum, cerebral cortex, medulla, spinal cord, occipital pole, frontal lobe, temporal pole, and putamen. *PSP1* mRNA was also detected in Alzheimer's disease brain.

Example 4 - Detecting the PSP1 polymorphisms

PSP1 oligonucleotides 1AFC, 1AFT and 1AR were designed for detecting the polymorphism at nucleotide 672 (cytidine to thymine) causing the Arg to Cys amino acid change. The Allele Specific Oligonucleotides (ASO) 1AFC and 1AFT are identical apart from their 3' end bases and provide the specificity for screening for the polymorphism.

1AFC: CAT CCG GCA TTG TTA GCT CTG C 22mer (SEQ ID NO:32)

1AFT: CAT CCG GCA TTG TTA GCT CTG T 22mer (SEQ ID NO:33)

1AR: CAA TAG CTG CAT CAG TTT GAA TG 23mer (SEQ ID NO:34)

Pairs of oligonucleotides (1AFC + 1AR, or 1AFT + 1AR) were used in a PCR under the following conditions: 94°C for 40 seconds, 60°C for 30 seconds, for 35 cycles in a reaction containing 1 U KlenTaq1 (GenPak Ltd.), 50mM Tris-Cl pH9.1, 16mM ammonium sulphate, 3.5mM $MgCl_2$, 150ug ml^{-1} BSA and 25ng of human genomic DNA of unknown source. Each pair of oligonucleotides was tested against 12 random samples of genomic DNA and the products electrophoresed on a 4% agarose (Gibco-BRL) gel. The expected product of 95 base pairs was seen for both ASOs in 8 of the 12 DNAs indicating that these individuals are heterozygous for this polymorphism. Two of the DNAs amplified with only the 1AFC oligonucleotide and are thus homozygous for the allele with the cytidine at this position. Two of the DNAs amplified with only the 1AFT oligonucleotide and are thus homozygous for the allele with the thymine at this position.

PSP1 oligonucleotides 1BFC, 1BFT and 1BR were designed for detecting the polymorphism at nucleotide 1435 (cytidine to thymine) causing the Ala to Val amino acid change.

1BFC: TGG CGG GCT TTG GGG GGC ATT C 22mer (SEQ ID NO:35)

1BFT: TGG CGG GCT TTG GGG GGC ATT T 22mer (SEQ ID NO:36)

1BR: GAC GTC AGC AGG GCC CGG AGG TC 23mer (SEQ ID NO:37)

Pairs of oligonucleotides (1BFC + 1BR, or 1BFT + 1BR) were used in a PCR under the following conditions:94°C for 40 seconds, 67°C for 30 seconds, for 35 cycles in a reaction containing 1 U KlenTaq1 (GenPak Ltd.), 50mM Tris-Cl pH9.1, 16mM ammonium sulphate, 3.5mM $MgCl_2$, 150ug ml^{-1} BSA and 25ng of human genomic DNA of unknown source. Each pair of oligonucleotides was tested against 12 random samples of genomic DNA and the products electrophoresed on a 4% agarose (Gibco-BRL) gel. The expected product of 75 base pairs was seen using the 1BFT ASO in 9 of the 12 samples indicating that the other 3 individuals have a different allele at this position.

Example 5 - Detecting the D87258 polymorphism

Oligonucleotides 2AFG, 2AFT and 2AR were designed for detecting the polymorphism at nucleotide 1325 (guanine to thymine) causing the Gly to Val amino acid change.

2AFG: GAT ACC CCA GCA GAA GCT GG 20mer (SEQ ID NO:38)

2AFT: GAT ACC CCA GCA GAA GCT GT 20mer (SEQ ID NO:39)

2AR: GCT GAC ATC ATT GGC GGA GAC 21mer (SEQ ID NO:40)

Pairs of oligonucleotides (2AFG + 2AR, or 2AFT + 2AR) were used in a PCR under the following conditions: 94°C for 40 seconds, 62°C for 30 seconds, for 35 cycles in a reaction containing 1 U KlenTaq1 (GenPak Ltd.), 50mM Tris-Cl pH9.1, 16mM ammonium sulphate, 3.5mM $MgCl_2$, 150ug ml^{-1} BSA and 25ng of human genomic DNA of unknown source. Each pair of oligonucleotides was tested against 12 random samples of genomic DNA and the products electrophoresed on a 4% agarose (Gibco-BRL) gel. The 2AFT ASO generated a band of approximately 1000 bp. The predicted band was 90 bp. Presumably, the presence of the larger bands was due to the presence of an intron in the region flanked by oligonucletides 2AR and 2AFT. Bands were observed in all of the samples amplified with 2AFT indicating that the allele containing the thymine is present in all 12 individuals.

The present invention may be embodied in other specific forms without departing from the spirit or essential attributes thereof, and, accordingly, reference should be made to the appended claims, rather than to the foregoing specification, as indicating the scope of the invention.

SEQUENCE LISTING

(1) GENERAL INFORMATION

(i) APPLICANT: Creasy, Caretha
Livi, George
Karran, Eric
Clinkenbeard, Helen
Browne, Michael
Southan, Christopher

(ii) TITLE OF THE INVENTION: HUMAN SERINE PROTEASE

(iii) NUMBER OF SEQUENCES: 40

(iv) CORRESPONDENCE ADDRESS:
(A) ADDRESSEE: SmithKline Beecham Corporation
(B) STREET: 709 Swedeland Road
(C) CITY: King of Prussia
(D) STATE: PA
(E) COUNTRY: USA
(F) ZIP: 19406

(v) COMPUTER READABLE FORM:
(A) MEDIUM TYPE: Diskette
(B) COMPUTER: IBM Compatible
(C) OPERATING SYSTEM: DOS
(D) SOFTWARE: FastSEQ Version 1.5

(vi) CURRENT APPLICATION DATA:
(A) APPLICATION NUMBER:
(B) FILING DATE:
(C) CLASSIFICATION:

(vii) PRIOR APPLICATION DATA:
(A) APPLICATION NUMBER: 60/025436
(B) FILING DATE: 06-SEPT-1996

(viii) ATTORNEY/AGENT INFORMATION:
(A) NAME: Baumeister, Kirk
(B) REGISTRATION NUMBER: 33,833
(C) REFERENCE/DOCKET NUMBER: P50547P2

(ix) TELECOMMUNICATION INFORMATION:
(A) TELEPHONE: 610-270-5096
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(C) TELEX:

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 732 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

```
GGGACTCCCC CAAACCAATG TGGAATACAT TCAAACTGAT GCAGCTATTG ATTTTGGAAA     60
CTCTGGAGGT CCCCTGGTTA ACCTGGATGG GGAGGTGATT GGAGTGAACA CCATGAAGGT    120
CACAGCTGGA ATCTCCTTTG CCATCCCTTC TGATCGTCTT CGAGAGTTTC TGCATCGTGG    180
GGAAAAGAAG AATTCCTCCT CCGGAATCAG TGGGTCCCAG CGGCGCTACA TTGGGGTGAT    240
GATGCTGACC CTGAGTCCCA GCATCCTTGC TGAACTACAG CTTCGAGAAC CAAGCTTTCC    300
CGATGTTCAG CATGGTGTAC TCATCCATAA AGTCATCCTG GGCTCCCCTG CACACCGGGC    360
TGGTCTGCGG CCTGGTGATG TGATTTTGGC CATTGGGGAG CAGATGGTAC AAAATGCTGA    420
AGATGTTTAT GAAGCTGTTC GAACCCAATC CCAGTTGGCA GTGCAGATCC GGCGGGGACG    480
AGAAACACTG ACCTTATATG TGACCCCTGA GGTCACAGAA TGAATAGATC ACCAAGAGTA    540
TGAGGCTCCT GCTCTGATTT CCTCCTTGCC TTTCTGGCTG AGGTTCTGAG GGCACCGAGA    600
CAGAGGGTTA AATGAACCAG TGGGGGCAGG TCCCTCCAAC CACCAGCACT GACTCCTGGG    660
CTCTGAAGAA TCACAGAAAC ACTTTTTATA TAAAATAAAA TTATACCTAG CAACAAAAAA    720
AAAAAAAAAA AA                                                        732
```

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 173 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE: N-terminal
(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

```
Gly Leu Pro Gln Thr Asn Val Glu Tyr Ile Gln Thr Asp Ala Ala Ile
 1               5                  10                  15
Asp Phe Gly Asn Ser Gly Gly Pro Leu Val Asn Leu Asp Gly Glu Val
            20                  25                  30
Ile Gly Val Asn Thr Met Lys Val Thr Ala Gly Ile Ser Phe Ala Ile
        35                  40                  45
Pro Ser Asp Arg Leu Arg Glu Phe Leu His Arg Gly Glu Lys Lys Asn
    50                  55                  60
Ser Ser Ser Gly Ile Ser Gly Ser Gln Arg Arg Tyr Ile Gly Val Met
65                  70                  75                  80
Met Leu Thr Leu Ser Pro Ser Ile Leu Ala Glu Leu Gln Leu Arg Glu
                85                  90                  95
Pro Ser Phe Pro Asp Val Gln His Gly Val Leu Ile His Lys Val Ile
            100                 105                 110
Leu Gly Ser Pro Ala His Arg Ala Gly Leu Arg Pro Gly Asp Val Ile
        115                 120                 125
Leu Ala Ile Gly Glu Gln Met Val Gln Asn Ala Glu Asp Val Tyr Glu
    130                 135                 140
Ala Val Arg Thr Gln Ser Gln Leu Ala Val Gln Ile Arg Arg Gly Arg
145                 150                 155                 160
Glu Thr Leu Thr Leu Tyr Val Thr Pro Glu Val Thr Glu
                165                 170
```

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1787 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

```
CCCAGTCTCT CGGCCCGGTT GTCTGTTGGG GTCACTGAAC CCCGAGCATG CCTGACGTCT     60
GGGACCCCGG GTCCCCGGGC ACAACTGACT GCGGTGACCC CAGATACCAG GACCCGGGAG    120
GCCTCAGAGA ACTCTGGAAC CCGTTCGCGC GCGTGGCTGG CGGTGGCGCT GGGCGCTGGG    180
GGGGCAGTGC TGTTGTTGTT GTGGGGCGGG GGTCGGGGTC CTCCGGCCGT CCTCGCCGCC    240
GTCCCTAGCC CGCCGCCCGC TTCTCCCCGG AGTCAGTACA ACTTCATCGC AGATGTGGTG    300
GAGAAGACAG CACCTGCCGT GGTCTATATC GAGATCCTGG ACCGGCACCC TTTCTTGGGC    360
CCCGAGGTCC CTATCTCGAA CGGCTCAGGA TTCGTGGTGG CTGCCGATGG GCTCATTGTC    420
ACCAACGCCC ATGTGGTGGC TGATCGGCGC AGAGTCCGTG TGAGACTGCT AAGCGGCGAC    480
ACGTATGAGG CCGTGGTCAC AGCTGTGGAT CCCGTGGCAG ACATCGCAAC GCTGAGGATT    540
CAGACTAAGG AGCCTCTCCC CACGCTGCCT CTGGGACGCT CAGCTGATGT CCGGCAAGGG    600
GAGTTTGTTG TTGCCATGGG AAGTCCCTTT GCACTGCAGA ACACGATCAC ATCCGGCATT    660
GTTAGCTCTG CTCAGCGTCC AGCCAGAGAC CTGGGACTCC CCCAAACCAA TGTGGAATAC    720
ATTCAAACTG ATGCAGCTAT TGATTTTGGA AACTCTGGAG GTCCCCTGGT TAACCTGGTG    780
AGTGAGACAT CCTTCCTTCC AAGAATCCCT GCCCCAGGTC AGTGTGGGAA GGGTAGGTTT    840
CCCCTAATTC AAGGATGTTT GGTCAAGTTT CTGAGCAGTT CTTTGTTGGC TATCTCTCAA    900
TATCCAACCA GATCTCCCCA ACACTTGCTG GTACTTTTGT TCGGGTGCCC CCATCCCCTA    960
CTATTTGTTT AGGCTAGGGA ACTGGGGGCT GTATCCCTGC AGGATGGGGA GGTGATTGGA   1020
GTGAACACCA TGAAGGTCAC AGCTGGAATC TCCTTTGCCA TCCCTTCTGA TCGTCTTCGA   1080
GAGTTTCTGC ATCGTGGGGA AAAGAACAAT TCCTCCTCCG GAATCAGTGG GTCCCAGCGG   1140
CGCTACATTG GGGTGATGAT GCTGACCCTG AGTCCCAGCA TCCTTGCTGA ACTACAGCTT   1200
CGAGAACCAA GCTTTCCCGA TGTTCAGCAT GGTGTACTCA TCCATAAAGT CATCCTGGGC   1260
TCCCCTGCAC ACCGGGCTGG TCTGCGGCCT GGTGATGTGA TTTTGGCCAT TGGGGAGCAG   1320
ATGGTACAAA ATGCTGAAGA TGTTTATGAA GCTGTTCGAA CCCAATCCCA GTTGGCAGTG   1380
CAGATCCGGC GGGGACGAGA AACACTGACC TTATATGTGA CCCCTGAGGT CACAGAATGA   1440
ATAGATCACC AAGAGTATGA GGCTCCTGCT CTGATTTCCT CCTTGCCTTT CTGGCTGAGG   1500
TTCTGAGGGC ACCGAGACAG AGGGTTAAAT GAACCAGTGG GGGCAGGTCC CTCCAACCAC   1560
CAGCACTGAC TCCTGGGCTC TGAAGAATCA CAGAAACACT TTTTATATAA AATAAAATTA   1620
TACCTAGCAA CATATTATAG TAAAAAATGA GGTGGGAGGG CTGGATCTTT TCCCCCACCA   1680
AAAGGCTAGA GGTAAAGCTG TATCCCCCTA AACTTAGGGG AGATACTGGA GCTGACCATC   1740
CTGACCTCCT ATTAAAGAAA ATGAGCTGCT GAAAAAAAAA AAAAAAA                 1787
```

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 323 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE: N-terminal
(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

```
Pro Ser Leu Trp Ala Arg Leu Ser Val Gly Val Thr Glu Pro Arg Ala
 1               5                  10                  15
Cys Leu Thr Ser Gly Thr Pro Gly Pro Arg Ala Gln Leu Thr Ala Val
            20                  25                  30
Thr Pro Asp Thr Arg Thr Arg Glu Ala Ser Glu Asn Ser Gly Thr Arg
        35                  40                  45
Ser Arg Ala Trp Leu Ala Val Ala Leu Gly Ala Gly Gly Ala Val Leu
    50                  55                  60
Leu Leu Leu Trp Gly Gly Gly Arg Gly Pro Pro Ala Val Leu Ala Ala
65                  70                  75                  80
Val Pro Ser Pro Pro Pro Ala Ser Pro Arg Ser Gln Tyr Asn Phe Ile
                85                  90                  95
Ala Asp Val Val Glu Lys Thr Ala Pro Ala Val Val Tyr Ile Glu Ile
            100                 105                 110
Leu Asp Arg His Pro Phe Leu Gly Arg Glu Val Pro Ile Ser Asn Gly
```

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        115                 120                 125
Ser Gly Phe Val Val Ala Ala Asp Gly Leu Ile Val Thr Asn Ala His
    130                 135                 140
Val Val Ala Asp Arg Arg Arg Val Arg Val Arg Leu Leu Ser Gly Asp
145                 150                 155                 160
Thr Tyr Glu Ala Val Val Thr Ala Val Asp Pro Val Ala Asp Ile Ala
                165                 170                 175
Thr Leu Arg Ile Gln Thr Lys Glu Pro Leu Pro Thr Leu Pro Leu Gly
            180                 185                 190
Arg Ser Ala Asp Val Arg Gln Gly Glu Phe Val Val Ala Met Gly Ser
        195                 200                 205
Pro Phe Ala Leu Gln Asn Thr Ile Thr Ser Gly Ile Val Ser Ser Ala
    210                 215                 220
Gln Arg Pro Ala Arg Asp Leu Gly Leu Pro Gln Thr Asn Val Glu Tyr
225                 230                 235                 240
Ile Gln Thr Asp Ala Ala Ile Asp Phe Gly Asn Ser Gly Gly Pro Leu
                245                 250                 255
Val Asn Leu Val Ser Glu Thr Ser Phe Leu Pro Arg Ile Pro Ala Pro
            260                 265                 270
Gly Gln Cys Gly Lys Gly Arg Phe Pro Leu Ile Gln Gly Cys Leu Val
        275                 280                 285
Lys Phe Leu Ser Ser Ser Leu Leu Ala Ile Ser Gln Tyr Pro Thr Arg
    290                 295                 300
Ser Pro Gln His Leu Leu Val Leu Leu Phe Gly Cys Pro His Pro Leu
305                 310                 315                 320
Leu Phe Val
```

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1503 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

```
CTTCGGGCAT GGCGGGCTTT GGGGGGCATT CGCTGGGGGA GGAGACCCCG TTTGACCCCT     60
GACCTCCGGG CCCTGCTGAC GTCAGGAACT TCTGACCCCC GGGCCCGAGT GACTTATGGG    120
ACCCCCAGTC TCTGGGCCCG GTTGTCTGTT GGGGTCACTG AACCCCGAGC ATGCCTGACG    180
TCTGGGACCC CGGGTCCCCG GGCACAACTG ACTGCGGTGA CCCCAGATAC CAGGACCCGG    240
GAGGCCTCAG AGAACTCTGG AACCCGTTCG CGCGCGTGGC TGGCCGTCGC GCTGGGCGCT    300
GGGGGGGCAG TGCTGTTGTT GTTGTGGGGC GGGGGTCGGG GTCCTCCGGC CGTCCTCGCC    360
GCCGTCCCTA GCCCGCCGCC CGCTTCTCCC CGGAGTCAGT ACAACTTCAT CGCAGATGTG    420
GTGGAGAAGA CAGCACCTGC CGTGGTCTAT ATCGAGATCC TGGACCGGCA CCCTTTCTTG    480
GGCCGCGAGG TCCCTATCTC GAACGGCTCA GGATTCGTGG TGGCTGCCGA TGGGCTCATT    540
GTCACCAACG CCCATGTGGT GGCTGATCGG CGCAGAGTCC GTGTGAGACT GCTAAGCGGC    600
GACACGTATG AGGCCGTGGT CACAGCTGTG GATCCCGTGG CAGACATCGC AACGCTGAGG    660
ATTCAGACTA AGGAGCCTCT CCCCACGCTG CCTCTGGGAC GCTCAGCTGA TGTCCGGCAA    720
GGGGAGTTTG TTGTTGCCAT GGGAAGTCCC TTTGCACTGC AGAACACGAT CACATCCGGC    780
ATTGTTAGCT CTGCTCAGCG TCCAGCCAGA GACCTGGGAC TCCCCCAAAC CAATGTGGAA    840
TACATTCAAA CTGATGCAGC TATTGATTTT GGAAACTCTG GAGGTCCCCT GGTTAACCTG    900
GCTAGGGAAC TGGGGGCTGT ATCCCTGCAG GATGGGGAGG TGATTGGAGT GAACACCATG    960
AAGGTCACAG CTGGAATCTC CTTTGCCATC CCTTCTGATC GTCTTCGAGA GTTTCTGCAT   1020
CGTGGGGAAA AGAAGAATTC CTCCTCCGGA ATCAGTGGGT CCCAGCGGCG CTACATTGGG   1080
GTCATGATGC TGACCCTGAG TCCCAGGGCT GGTCTGCGGC CTGGTGATGT GATTTTGGCC   1140
ATTGGGGAGC AGATGGTACA AAATGCTGAA GATGTTTATG AAGCTGTTCG AACCCAATCC   1200
CAGTTGGCAG TGCAGATCCG GCGGGGACGA GAAACACTGA CCTTATATGT GACCCCTGAG   1260
```

```
GTCACAGAAT GAATAGATCA CCAAGAGTAT GAGGCTCCTG CTCTGATTTC CTCCTTGCCT   1320
TTCTGGCTGA GGTTCTGAGG GCACCGAGAC AGACGGTTAA ATGAACCAGT GGGGGCAGGT   1380
CCCTCCAACC ACCAGCACTG ACTCCTGGGC TCTGAAGAAT CACAGAAACA CTTTTTATAT   1440
AAAATAAAAT TATACCTAGC AAAAAAAAAA AAAAAAAAAA AAAAAAAAAA AAAAAAAAAA   1500
AAA                                                                 1503
```

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 423 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE: N-terminal
(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

```
Leu Arg Ala Trp Arg Ala Leu Gly Gly Ile Arg Trp Gly Arg Arg Pro
 1               5                  10                  15
Arg Leu Thr Pro Asp Leu Arg Ala Leu Leu Thr Ser Gly Thr Ser Asp
            20                  25                  30
Pro Arg Ala Arg Val Thr Tyr Gly Thr Pro Ser Leu Trp Ala Arg Leu
        35                  40                  45
Ser Val Gly Val Thr Glu Pro Arg Ala Cys Leu Thr Ser Gly Thr Pro
    50                  55                  60
Gly Pro Arg Ala Gln Leu Thr Ala Val Thr Pro Asp Thr Arg Thr Arg
65                  70                  75                  80
Glu Ala Ser Glu Asn Ser Gly Thr Arg Ser Arg Ala Trp Leu Ala Val
                85                  90                  95
Ala Leu Gly Ala Gly Gly Ala Val Leu Leu Leu Leu Trp Gly Gly Gly
            100                 105                 110
Arg Gly Pro Pro Ala Val Leu Ala Ala Val Pro Ser Pro Pro Pro Ala
        115                 120                 125
Ser Pro Arg Ser Gln Tyr Asn Phe Ile Ala Asp Val Val Glu Lys Thr
    130                 135                 140
Ala Pro Ala Val Val Tyr Ile Glu Ile Leu Asp Arg His Pro Phe Leu
145                 150                 155                 160
Gly Arg Glu Val Pro Ile Ser Asn Gly Ser Gly Phe Val Val Ala Ala
                165                 170                 175
Asp Gly Leu Ile Val Thr Asn Ala His Val Val Ala Asp Arg Arg Arg
            180                 185                 190
Val Arg Val Arg Leu Leu Ser Gly Asp Thr Tyr Glu Ala Val Val Thr
        195                 200                 205
Ala Val Asp Pro Val Ala Asp Ile Ala Thr Leu Arg Ile Gln Thr Lys
    210                 215                 220
Glu Pro Leu Pro Thr Leu Pro Leu Gly Arg Ser Ala Asp Val Arg Gln
225                 230                 235                 240
Gly Glu Phe Val Val Ala Met Gly Ser Pro Phe Ala Leu Gln Asn Thr
                245                 250                 255
Ile Thr Ser Gly Ile Val Ser Ser Ala Gln Arg Pro Ala Arg Asp Leu
            260                 265                 270
Gly Leu Pro Gln Thr Asn Val Glu Tyr Ile Gln Thr Asp Ala Ala Ile
        275                 280                 285
Asp Phe Gly Asn Ser Gly Gly Pro Leu Val Asn Leu Ala Arg Glu Leu
    290                 295                 300
Gly Ala Val Ser Leu Gln Asp Gly Glu Val Ile Gly Val Asn Thr Met
305                 310                 315                 320
Lys Val Thr Ala Gly Ile Ser Phe Ala Ile Pro Ser Asp Arg Leu Arg
                325                 330                 335
Glu Phe Leu His Arg Gly Glu Lys Lys Asn Ser Ser Ser Gly Ile Ser
```

```
            340                 345                 350
Gly Ser Gln Arg Arg Tyr Ile Gly Val Met Met Leu Thr Leu Ser Pro
        355                 360                 365
Arg Ala Gly Leu Arg Pro Gly Asp Val Ile Leu Ala Ile Gly Glu Gln
    370                 375                 380
Met Val Gln Asn Ala Glu Asp Val Tyr Glu Ala Val Arg Thr Gln Ser
385                 390                 395                 400
Gln Leu Ala Val Gln Ile Arg Arg Gly Arg Glu Thr Leu Thr Leu Tyr
                405                 410                 415
Val Thr Pro Glu Val Thr Glu
            420
```

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1835 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:
(ix) FEATURE:

(A) NAME/KEY: Coding Sequence
(B) LOCATION: 251...1624
(D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

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GGCCGGAAGG GCTAGCGGTC CCAGCATACC CCGCGGCCCC TTGGGCCGTC TCACAACTCG     60
CGTCCGGCGG AGACCACAAT TCCCGGCATT CGTGGGGCAT GGAGGAGTCG GCCTCCCGGA    120
ATCCTGGTCC CGGCGTGCAC TTCTGAAGGA CTTCAGGTAC CGGCGTGCCC CGCGTCCTAC    180
TGTCCGCCTG CTCGCGTCCT GGGTGCCGCC TCTGAGTAGG GCGGGCGAGG AGGCAGCCAA    240
GGCGGAGCTG  ATG GCT GCC CCG AGG GCG GGG CGG GGT GCA GGC TGG AGC       289
           Met Ala Ala Pro Arg Ala Gly Arg Gly Ala Gly Trp Ser
            1               5                  10
CTT CGG GCA TGG CGG GCT TTG GGG GGC ATT TGC TGG GGG AGG AGA CCC      337
Leu Arg Ala Trp Arg Ala Leu Gly Gly Ile Cys Trp Gly Arg Arg Pro
    15                  20                  25

CGT TTG ACC CCT GAC CTC CGG GCC CTG CTG ACG TCA GGA ACT TCT GAC      385
Arg Leu Thr Pro Asp Leu Arg Ala Leu Leu Thr Ser Gly Thr Ser Asp
30                  35                  40                  45

CCC CGG GCC CGA GTG ACT TAT GGG ACC CCC AGT CTC TGG GCC CGG TTG      433
Pro Arg Ala Arg Val Thr Tyr Gly Thr Pro Ser Leu Trp Ala Arg Leu
                50                  55                  60

TCT GTT GGG GTC ACT GAA CCC CGA GCA TGC CTG ACG TCT GGG ACC CCG      481
Ser Val Gly Val Thr Glu Pro Arg Ala Cys Leu Thr Ser Gly Thr Pro
            65                  70                  75

GGT CCC CGG GCA CAA CTG ACT GCG GTG ACC CCA GAT ACC AGG ACC CGG      529
Gly Pro Arg Ala Gln Leu Thr Ala Val Thr Pro Asp Thr Arg Thr Arg
        80                  85                  90

GAG GCC TCA GAG AAC TCT GGA ACC CGT TCG CGC GCG TGG CTG GCG GTG      577
Glu Ala Ser Glu Asn Ser Gly Thr Arg Ser Arg Ala Trp Leu Ala Val
    95                  100                 105
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GCG CTG GGC GCT GGG GGG GCA GTG CTG TTG TTG TTG TGG GGC GGG GGT      625
Ala Leu Gly Ala Gly Gly Ala Val Leu Leu Leu Leu Trp Gly Gly Gly
110                 115                 120                 125

CGG GGT CCT CCG GCC GTC CTC GCC GCC GTC CCT AGC CCG CCG CCC GCT      673
Arg Gly Pro Pro Ala Val Leu Ala Ala Val Pro Ser Pro Pro Pro Ala
                130                 135                 140

TCT CCC CGG AGT CAG TAC AAC TTC ATC GCA GAT GTG GTG GAG AAG ACA      721
Ser Pro Arg Ser Gln Tyr Asn Phe Ile Ala Asp Val Val Glu Lys Thr
            145                 150                 155

GCA CCT GCC GTG GTC TAT ATC GAG ATC CTG GAC CGG CAC CCT TTC TTG      769
Ala Pro Ala Val Val Tyr Ile Glu Ile Leu Asp Arg His Pro Phe Leu
        160                 165                 170

GGC CGC GAG GTC CCT ATC TCG AAC GGC TCA GGA TTC GTG GTG GCT GCC      817
Gly Arg Glu Val Pro Ile Ser Asn Gly Ser Gly Phe Val Val Ala Ala
    175                 180                 185

GAT GGG CTC ATT GTC ACC AAC GCC CAT GTG GTG GCT GAT CGG CGC AGA      865
Asp Gly Leu Ile Val Thr Asn Ala His Val Val Ala Asp Arg Arg Arg
190                 195                 200                 205

GTC CGT GTG AGA CTG CTA AGC GGC GAC ACG TAT GAG GCC GTG GTC ACA      913
Val Arg Val Arg Leu Leu Ser Gly Asp Thr Tyr Glu Ala Val Val Thr
                210                 215                 220

GCT GTG GAT CCC GTG GCA GAC ATC GCA ACG CTG AGG ATT CAG ACT AAG      961
Ala Val Asp Pro Val Ala Asp Ile Ala Thr Leu Arg Ile Gln Thr Lys
            225                 230                 235

GAG CCT CTC CCC ACG CTG CCT CTG GGA CGC TCA GCT GAT GTC CGG CAA     1009
Glu Pro Leu Pro Thr Leu Pro Leu Gly Arg Ser Ala Asp Val Arg Gln
        240                 245                 250

GGG GAG TTT GTT GTT GCC ATG GGA AGT CCC TTT GCA CTG CAG AAC ACG     1057
Gly Glu Phe Val Val Ala Met Gly Ser Pro Phe Ala Leu Gln Asn Thr
    255                 260                 265

ATC ACA TCC GGC ATT GTT AGC TCT GCT CAG CGT CCA GCC AGA GAC CTG     1105
Ile Thr Ser Gly Ile Val Ser Ser Ala Gln Arg Pro Ala Arg Asp Leu
270                 275                 280                 285

GGA CTC CCC CAA ACC AAT GTG GAA TAC ATT CAA ACT GAT GCA GCT ATT     1153
Gly Leu Pro Gln Thr Asn Val Glu Tyr Ile Gln Thr Asp Ala Ala Ile
                290                 295                 300

GAT TTT GGA AAC TCT GGA GGT CCC CTG GTT AAC CTG GAT GGC GAG GTG     1201
Asp Phe Gly Asn Ser Gly Gly Pro Leu Val Asn Leu Asp Gly Glu Val
            305                 310                 315

ATT GGA GTG AAC ACC ATG AAG GTC ACA GCT GGA ATC TCC TTT GCC ATC     1249
Ile Gly Val Asn Thr Met Lys Val Thr Ala Gly Ile Ser Phe Ala Ile
        320                 325                 330

CCT TCT GAT CGT CTT CGA GAG TTT CTG CAT CGT GGG GAA AAG AAG AAT     1297
Pro Ser Asp Arg Leu Arg Glu Phe Leu His Arg Gly Glu Lys Lys Asn
    335                 340                 345

TCC TCC TCC GGA ATC AGT GGG TCC CAG CGG CGC TAC ATT GGG GTG ATG     1345
Ser Ser Ser Gly Ile Ser Gly Ser Gln Arg Arg Tyr Ile Gly Val Met
350                 355                 360                 365

ATG CTG ACC CTG AGT CCC AGC ATC CTT GCT GAA CTA CAG CTT CGA GAA     1393
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Met Leu Thr Leu Ser Pro Ser Ile Leu Ala Glu Leu Gln Leu Arg Glu
                370                 375                 380

CCA AGC TTT CCC GAT GTT CAG CAT GGT GTA CTC ATC CAT AAA GTC ATC    1441
Pro Ser Phe Pro Asp Val Gln His Gly Val Leu Ile His Lys Val Ile
            385                 390                 395

CTG GGC TCC CCT GCA CAC CGG GCT GGT CTG CGG CCT GGT GAT GTG ATT    1489
Leu Gly Ser Pro Ala His Arg Ala Gly Leu Arg Pro Gly Asp Val Ile
        400                 405                 410

TTG GCC ATT GGG GAG CAG ATG GTA CAA AAT GCT GAA GAT GTT TAT GAA    1537
Leu Ala Ile Gly Glu Gln Met Val Gln Asn Ala Glu Asp Val Tyr Glu
    415                 420                 425

GCT GTT CGA ACC CAA TCC CAG TTG GCA GTG CAG ATC CGG CGG GGA CGA    1585
Ala Val Arg Thr Gln Ser Gln Leu Ala Val Gln Ile Arg Arg Gly Arg
430                 435                 440                 445

GAA ACA CTG ACC TTA TAT GTG ACC CCT GAG GTC ACA GAA TGAATAGATC ACC  1637
Glu Thr Leu Thr Leu Tyr Val Thr Pro Glu Val Thr Glu
                450                 455

AAGAGTATGA GGCTCCTGCT CTGATTTCCT CCTTGCCTTT CTGGCTGAGG TTCTGAGGGC  1697
ACCGAGACAG AGGGTTAAAT GAACCAGTGG GGGCAGGTCC CTCCAACCAC CAGCACTGAC  1757
TCCTGGGCTC TGAAGAATCA CAGAAACACT TTTTATATAA AATAAAATTA TACCTAGCAA  1817
CATAAAAAAA AAAAAAAA                                                1835
```

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 458 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE: internal
(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

```
Met Ala Ala Pro Arg Ala Gly Arg Gly Ala Gly Trp Ser Leu Arg Ala
 1               5                  10                  15
Trp Arg Ala Leu Gly Gly Ile Cys Trp Gly Arg Arg Pro Arg Leu Thr
            20                  25                  30
Pro Asp Leu Arg Ala Leu Leu Thr Ser Gly Thr Ser Asp Pro Arg Ala
        35                  40                  45
Arg Val Thr Tyr Gly Thr Pro Ser Leu Trp Ala Arg Leu Ser Val Gly
    50                  55                  60
Val Thr Glu Pro Arg Ala Cys Leu Thr Ser Gly Thr Pro Gly Pro Arg
65                  70                  75                  80
Ala Gln Leu Thr Ala Val Thr Pro Asp Thr Arg Thr Arg Glu Ala Ser
                85                  90                  95
Glu Asn Ser Gly Thr Arg Ser Arg Ala Trp Leu Ala Val Ala Leu Gly
            100                 105                 110
Ala Gly Gly Ala Val Leu Leu Leu Leu Trp Gly Gly Gly Arg Gly Pro
        115                 120                 125
Pro Ala Val Leu Ala Ala Val Pro Ser Pro Pro Pro Ala Ser Pro Arg
    130                 135                 140
Ser Gln Tyr Asn Phe Ile Ala Asp Val Val Glu Lys Thr Ala Pro Ala
145                 150                 155                 160
Val Val Tyr Ile Glu Ile Leu Asp Arg His Pro Phe Leu Gly Arg Glu
```

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                165                 170                 175
Val Pro Ile Ser Asn Gly Ser Gly Phe Val Val Ala Ala Asp Gly Leu
            180                 185                 190
Ile Val Thr Asn Ala His Val Val Ala Asp Arg Arg Arg Val Arg Val
        195                 200                 205
Arg Leu Leu Ser Gly Asp Thr Tyr Glu Ala Val Val Thr Ala Val Asp
    210                 215                 220
Pro Val Ala Asp Ile Ala Thr Leu Arg Ile Gln Thr Lys Glu Pro Leu
225                 230                 235                 240
Pro Thr Leu Pro Leu Gly Arg Ser Ala Asp Val Arg Gln Gly Glu Phe
                245                 250                 255
Val Val Ala Met Gly Ser Pro Phe Ala Leu Gln Asn Thr Ile Thr Ser
            260                 265                 270
Gly Ile Val Ser Ser Ala Gln Arg Pro Ala Arg Asp Leu Gly Leu Pro
        275                 280                 285
Gln Thr Asn Val Glu Tyr Ile Gln Thr Asp Ala Ala Ile Asp Phe Gly
    290                 295                 300
Asn Ser Gly Gly Pro Leu Val Asn Leu Asp Gly Glu Val Ile Gly Val
305                 310                 315                 320
Asn Thr Met Lys Val Thr Ala Gly Ile Ser Phe Ala Ile Pro Ser Asp
                325                 330                 335
Arg Leu Arg Glu Phe Leu His Arg Gly Glu Lys Lys Asn Ser Ser Ser
            340                 345                 350
Gly Ile Ser Gly Ser Gln Arg Arg Tyr Ile Gly Val Met Met Leu Thr
        355                 360                 365
Leu Ser Pro Ser Ile Leu Ala Glu Leu Gln Leu Arg Glu Pro Ser Phe
    370                 375                 380
Pro Asp Val Gln His Gly Val Leu Ile His Lys Val Ile Leu Gly Ser
385                 390                 395                 400
Pro Ala His Arg Ala Gly Leu Arg Pro Gly Asp Val Ile Leu Ala Ile
                405                 410                 415
Gly Glu Gln Met Val Gln Asn Ala Glu Asp Val Tyr Glu Ala Val Arg
            420                 425                 430
Thr Gln Ser Gln Leu Ala Val Gln Ile Arg Arg Gly Arg Glu Thr Leu
        435                 440                 445
Thr Leu Tyr Val Thr Pro Glu Val Thr Glu
    450                 455
```

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 2764 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

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TGGGACAGGC AGCTCCGGGG TCCGCGGTTT CACATCGGAA ACAAAACAGC GGCTGGTCTG     60
GAAGGAACCT GAGCTACGAG CCGCGGCGGC AGCGGGGCGG CGGGGAAGCG TATACCTAAT    120
CTGGGAGCCT GCAAGTGACA ACAGCCTTTG CGGTCCTTAG ACAGCTTGGC CTGGAGGAGA    180
ACACATGAAA GAAAGAACCT CAAGAGGCTT TGTTTTCTGT GAAACAGTAT TTCTATACAG    240
TTGCTCCAAT GACAGAGTTA CCTGCACCGT TGTCCTACTT CCAGAATGCA CAGATGTCTG    300
AGGACAACCA CCTGAGCAAT ACTGTACGTA GCCAGAATGA CAATAGAGAA CGGCAGGAGC    360
ACAACGACAG ACGGAGCCTT GGCCACCCTG AGCCATTATC TAATGGACGA CCCCAGGGTA    420
ACTCCCGGCA GGTGGTGGAG CAAGATGAGG AAGAAGATGA GGAGCTGACA TTGAAATATG    480
GCGCCAAGCA TGTGATCATG CTCTTTGTCC CTGTGACTCT CTGCATGGTG GTGGTCGTGG    540
CTACCATTAA GTCAGTCAGC TTTTATACCC GGAAGGATGG GCAGCTAATC TATACCCCAT    600
TCACAGAAGA TACCGAGACT GTGGGCCAGA GAGCCCTGCA CTCAATTCTG AATGCTGCCA    660
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TCATGATCAG TGTCATTGTT GTCATGACTA TCCTCCTGGT GGTTCTGTAT AAATACAGGT    720
GCTATAAGGT CATCCATGCC TGGCTTATTA TATCATCTCT ATTGTTGCTG TTCTTTTTTT    780
CATTCATTTA CTTGGGGGAA GTGTTTAAAA CCTATAACGT TGCTGTGGAC TACATTACTG    840
TTGCACTCCT GATCTGGAAT TTTGGTGTGG TGGGAATGAT TTCCATTCAC TGGAAAGGTC    900
CACTTCGACT CCAGCAGGCA TATCTCATTA TGATTAGTGC CCTCATGGCC CTGGTGTTTA    960
TCAAGTACCT CCCTGAATGG ACTGCGTGGC TCATCTTGGC TGTGATTTCA GTATATGATT   1020
TAGTGGCTGT TTTGTGTCCG AAAGGTCCAC TTCGTATGCT GGTTGAAACA GCTCAGGAGA   1080
GAAATGAAAC GCTTTTTCCA GCTCTCATTT ACTCCTCAAC AATGGTGTGG TTGGTGAATA   1140
TGGCAGAAGG AGACCCGGAA GCTCAAAGGA GAGTATCCAA AAATTCCAAG TATAATGCAG   1200
AAAGCACAGA AAGGGAGTCA CAAGACACTG TTGCAGAGAA TGATGATGGC GGGTTCAGTG   1260
AGGAATGGGA AGCCCAGAGG GACAGTCATC TAGGGCCTCA TCGCTCTACA CCTGAGTCAC   1320
GAGCTGCTGT CCAGGAACTT TCCAGCAGTA TCCTCGCTGG TGAAGACCCA GAGGAAAGGG   1380
GAGTAAAACT TGGATTGGGA GATTTCATTT TCTACAGTGT TCTGGTTGGT AAAGCCTCAG   1440
CAACAGCCAG TGGAGACTGG AACACAACCA TAGCCTGTTT CGTAGCCATA TTAATTGGTT   1500
TGTGCCTTAC ATTATTACTC CTTGCCATTT TCAAGAAAGC ATTGCCAGCT CTTCCAATCT   1560
CCATCACCTT TGGGCTTGTT TTCTACTTTG CCACAGATTA TCTTGTACAG CCTTTTATGG   1620
ACCAATTAGC ATTCCATCAA TTTTATATCT AGCATATTTG CGGTTAGAAT CCCATGGATG   1680
TTTCTTCTTT GACTATAACC AAATCTGGGG AGGACAAAGG TGATTTTCCT GTGTCCACAT   1740
CTAACAAAGT CAAGATTCCC GGCTGGACTT TTGCAGCTTC CTTCCAAGTC TTCCTGACCA   1800
CCTTGCACTA TTGGACTTTG GAAGGAGGTG CCTATAGAAA ACGATTTTGA ACATACTTCA   1860
TCGCAGTGGA CTGTGTCCCT CGGTGCAGAA ACTACCAGAT TTGAGGGACG AGGTCAAGGA   1920
GATATCATAG GCCCGGAAGT TGCTGTGCCC CATCAGCAGC TTGACGCGTG GTCACAGGAC   1980
GATTTCACTG ACACTGCGAA CTCTCAGGAC TACCGGTTAC CAAGAGGTTA GGTGAAGTGG   2040
TTTAAACCAA ACGGAACTCT TCATCTTAAA CTACACGTTG AAAATCAACC CAATAATTCT   2100
GTATTAACTG AATTCTGAAC TTTTCAGGAG GTACTGTGAG GAAGAGCAGG CACCAGCAGC   2160
AGAATGGGGA ATGGAGAGGT GGGCAGGGGT TCCAGCTTCC CTTTGATTTT TTGCTGCAGA   2220
CTCATCCTTT TTAAATGAGA CTTGTTTTCC CCTCTCTTTG AGTCAAGTCA AATATGTAGA   2280
TTGCCTTTGG CAATTCTTCT TCTCAAGCAC TGACACTCAT TACCGTCTGT GATTGCCATT   2340
TCTTCCCAAG GCCAGTCTGA ACCTGAGGTT GCTTTATCCT AAAAGTTTTA ACCTCAGGTT   2400
CCAAATTCAG TAAATTTTGG AAACAGTACA GCTATTTCTC ATCAATTCTC TATCATGTTG   2460
AAGTCAAATT TGGATTTTCC ACCAAATTCT GAATTTGTAG ACATACTTGT ACGCTCACTT   2520
GCCCCCAGAT GCCTCCTCTG TCCTCATTCT TCTCTCCCAC ACAAGCAGTC TTTTTCTACA   2580
GCCAGTAAGG CAGCTCTGTC RTGGTAGCAG ATGGTCCCAT TATTCTAGGG TCTTACTCTT   2640
TGTATGATGA AAAGAATGTG TTATGAATCG GTGCTGTCAG CCCTGCTGTC AGACCTTCTT   2700
CCACAGCAAA TGAGATGTAT GCCCAAAGCG GTAGAATTAA AGAAGAGTAA AATGGCTGTT   2760
GAAG                                                               2764
```

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 467 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide
(iii) HYPOTHETICAL: NO
(iv) ANTISENSE: NO
(v) FRAGMENT TYPE: N-terminal
(vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

```
Met Thr Glu Leu Pro Ala Pro Leu Ser Tyr Phe Gln Asn Ala Gln Met
 1               5                  10                  15
Ser Glu Asp Asn His Leu Ser Asn Thr Val Arg Ser Gln Asn Asp Asn
            20                  25                  30
Arg Glu Arg Gln Glu His Asn Asp Arg Arg Ser Leu Gly His Pro Glu
        35                  40                  45
Pro Leu Ser Asn Gly Arg Pro Gln Gly Asn Ser Arg Gln Val Val Glu
    50                  55                  60
Gln Asp Glu Glu Glu Asp Glu Glu Leu Thr Leu Lys Tyr Gly Ala Lys
65                  70                  75                  80
His Val Ile Met Leu Phe Val Pro Val Thr Leu Cys Met Val Val Val
                85                  90                  95
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